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(54) Title: A GENE RELATED TO MIGRAINE IN MAN**(57) Abstract**

Genes for familial hemiplegic migraine (FHM), episodic ataxia type-2 (EA-2), common forms of migraine, and other episodic neurological disorders, such as epilepsy, have been mapped to chromosome 19p13 and chromosome 10p12. A brain-specific P/Q-type calcium channel subunit gene, covering 300 kb with 47 exons is provided. The exons and their surroundings reveal polymorphic variations and deleterious mutations that are linked to various types of calcium channel dysfunctions causing episodic neurological disorders in man or animals.

Title: A gene related to migraine in man.

Migraine is a frequent paroxysmal neuro-vascular disorder, characterized by recurrent attacks of disabling headache, vomiting, photo/phonophobia, malaise, and other general symptoms (migraine without aura). Up to 20% of
5 patients may, in addition, experience transient neurological (aura) symptoms during attacks (migraine with aura) (HCC, 1988). Up to 24% of females and 12% of males in the general population are affected, however with variable attack frequency, duration and severity (Russell et al., 1995).
10 Knowledge about the mechanisms of the final common pathway of migraine attacks has increased substantially the last five years, resulting in improved, though still only symptomatic (and sub-optimal) acute treatment for the attack. There is, however, still very little knowledge about the etiology of
15 migraine attacks, i.e. why and how attacks begin and recur. Accordingly, prophylactic treatment for migraine is non-specific and has only limited efficacy.

Family, twin and population-based studies suggest that genetic factors are involved in migraine, most likely as part
20 of a multifactorial mechanism (reviewed by Haan et al., 1996). The complex genetics has hampered identification of candidate genes for migraine. Familial Hemiplegic Migraine (FHM) is a rare, autosomal dominant, subtype of migraine with aura, associated with ictal hemiparesis and, in some families
25 cerebellar atrophy (HCC, 1988). Otherwise, the symptoms of the headache and aura phase of FHM and "normal" migraine attacks are very similar and both types of attacks may alternate within subject and co-occur within families. FHM is thus part of the migraine spectrum and can be used as a model
30 to study the complex genetics of the more common forms of migraine (Haan et al., 1996). A gene for FHM has been assigned to chromosome 19p13 in about half of the families tested (Joutel et al., 1993; Ophoff et al., 1994;

Joutel et al., 1995). Remarkably, cerebellar atrophy was found only in families with FHM linked to chromosome 19p13, but not in unlinked families. Recently, we showed the 19p13 FHM locus to be also involved in "normal" migraine (May et al., 1995).

Episodic ataxia type 2 (EA-2) is another, autosomal dominant, paroxysmal neurological disorder, characterized by acetazolamide-responsive attacks of cerebellar ataxia and migraine-like symptoms, and interictal nystagmus and cerebellar atrophy. Recently, a gene for EA-2 was assigned to chromosome 19p13, within the same interval as for FHM (Kramer et al., 1995). This finding, as well as the clinical similarities, raise the possibility of EA-2 and FHM being allelic disorders.

Since other hereditary episodic neurological disorders responding to acetazolamide (such as hypokalaemic and hyperkalaemic periodic paralysis), as well as EA type-1 (which, in contrast to EA-2, is associated with continuous myokymia and non-responsive to acetazolamide) have all been associated with mutations in genes encoding for ion channels (Ptacek et al., 1991; Ptacek et al., 1994; Brown et al., 1994), we specifically looked for similar genes within the FHM and EA-2 candidate region.

In view of the above, the FHM/EA-2 locus can, since FHM is part of the migraine spectrum, thus be used to study the genetic factors and biological mechanisms that are related to various episodic neurological disorders such as FHM, EA-2, common migraine and others such as epilepsy.

Calcium channels are multisubunit complexes composed of at least an $\alpha 1$, an $\alpha 2\delta$, and a β subunit. The central $\alpha 1$ subunit is functionally the most important component, acting as a voltage sensor and forming the ion-conducting pore. The other subunits have auxiliary regulatory roles. The membrane topology of the $\alpha 1$ subunit consist of four hydrophobic motifs (I to IV), each containing six transmembrane α -helices (S1-

S6) and one hairpin (P) between S5-S6 that spans only the outer part of the transmembrane region.

The present invention provides an isolated and/or recombinant nucleic acid, or fragments thereof, encoding a Ca²⁺-channel α 1 subunit related to familial hemiplegic migraine and/or episodic ataxia type-2, derived from a gene present on chromosome 19p13.1-19p13.2; a gene encoding the α 1 (ion-conducting) subunit of a P/Q-type voltage gated calcium channel. The present invention also provides access to and methods to study the genetic background and identify other subunits of the calcium channel subunit complexes and the proteins related therewith that are associated with the genetic factors and biological mechanisms that are related to various episodic neurological disorders such as FHM, EA-2, common migraine and others such as epilepsy which are related to cation channel dysfunction.

The sequence of the cDNA of the gene is highly related ($\geq 90\%$) to a brain-specific rabbit and rat voltage gated P/Q-type calcium channel α 1 subunit (Mori et al., 1991; Starr et al., 1991), and the open reading frame consists of 2261 amino acid residues. Northern blot analysis showed a brain-specific expression, especially in the cerebellum. Primary study of a cosmid contig harbouring the gene already indicated an exon distribution over at least 300 kb of genomic DNA. Recently, a neuronal Ca²⁺ α 1A subunit gene was localized to chromosome 19p13.1-p13.2 by FISH analysis (Diriong et al, 1995). The gene symbol is CACNL1A4 and the α 1 subunit is classified as a P/Q-type. No sequence data for the CACNL1A4 gene have been provided by Diriong or others, but the same localization (chromosome 19p13.1) and the identical classification (P/Q-type) suggests that the Ca²⁺ channel α 1 subunit we have identified is very similar to CACNL1A4. No relation with migraine has been disclosed for CACNL1A4. The genomic structures of three other human Ca²⁺ channel α 1 subunit genes (CACNL1A1, CACNL1A2 and CACNL1A3) have been published to date (Hogan et al, 1994; Soldatov, 1994; Yamada et al, 1995). Both CACNL1A1 and CACNL1A2 span

about 150 kb and consist of 50 and 49 exons, respectively. The smaller CACNL1A3 gene is composed of 44 exons, distributed over 90 kb.

The present invention also provides an isolated and/or recombinant nucleic acid comprising alleles of the invented gene which contain mutations relevant to the occurrence of migraine and other neurological disorders which are related to cation channel dysfunction. Such mutations are for example a mutation at codon 192 resulting in the replacement of arginine by glutamine (R192Q), and/or a mutation at codon 666 resulting in the replacement of threonine by methionine, and/or a mutation at codon 714 resulting in a replacement of valine by alanine and/or a mutation at codon 1811 resulting in a replacement of isoleucine by leucine, but also other mutations of alleles of said gene which bear relationships with cation channel dysfunction.

The present invention also provides isolated and/or recombinant nucleic acid comprising alleles of said gene which contain a polymorphic CA-repeat sequence specific for various alleles of said gene. The present invention also provides isolated and/or recombinant nucleic acids comprising alleles of said gene which contain a CAG repeat.

The present invention also provides methods and tests (such as PCR, but also other tests to detect or amplify nucleic acids are known in the art) to detect, identify and localize or distinguish genes and alleles of such genes, or fragments thereof, encoding for proteins or α , β or χ subunits of specific cerebral cation channels, more specifically the invented gene and its various alleles encoding the $\alpha 1$ subunit of a P/Q-type voltage gated calcium channel and the gene encoding the $\beta 2$ sub-unit, which are involved in the primary pathogenesis of neurological disorders such as FHM, migraine, EA-2 and SCA6. With such methods and tests one can study abnormalities of said gene.

The invention also provides recombinant expression vectors comprising isolated and/or recombinant nucleic acid comprising alleles of said genes or fragments thereof, and

provides host cells or animals that comprise such vectors or that are otherwise transformed with an isolated and/or recombinant nucleic acid according to the invention.

The invention thus also provides a rationale and methods
5 for the testing and the development of specific prophylactic medication for migraine and other episodic neurological, in particular brain, disorders, such as epilepsy, associated with cation channel dysfunction.

The invention for example provides cells or animals that
10 comprise recombinant vectors that comprise variants of said genes or cells or animals that are transformed with said variants. Also, the invention provides means to identify naturally occurring variants of experimental animals with changes in said gene related to FHM, EA-2, SCA7, migraine or
15 other neurological disorders associated with cation channel dysfunction. An example of such an animal is the tottering mouse, and its variants called leaner and rolling, described in the experimental part of the invention. The invention also provides cells or animals in which changes such as deletions
20 or mutations in said gene have been introduced by recombinant nucleic acid techniques. All such cells or animals provided by the invention can be used to study the pathophysiology of FHM, EA-2, migraine or other neurological disorders associated with cation channel dysfunction, for example to
25 test or develop specific medication for the treatment of said disorders.

The invention also provides proteins or peptides encoded by said genes, or fragments thereof, related with cation channel dysfunction, and detection of such proteins or
30 peptides by antibodies directed against said proteins or peptides. Such antibodies can be of natural or synthetic origin, and can be produced by methods known in the art. Such proteins and antibodies and detection methods can be used to further in vitro or in vivo studies towards the
35 pathophysiology of FHM, EA-2, migraine or other neurological disorders associated with cation channel dysfunction, in addition such proteins, antibodies and detection methods can

also be used to diagnose or identify such disorders in patients or in experimental animals.

Experimental Procedures

5

Subjects

Sixteen FHM patients were selected, including eight individuals from four unrelated chromosome 19-linked FHM families (*NL-A*, *UK-B*, *USA-C* (Ophoff et al, 1994), and *USA-P* (Elliot et al., 1995), two affected individuals from two small FHM families from Italy (*Italy I & II*) and six individuals with sporadic hemiplegic migraine (i.e. no other family member was shown to suffer from attacks of hemiplegic migraine). In families *NL-A* and *USA-P* cerebellar ataxia and/or nystagmus is associated with FHM. An additional set of four subjects from four unrelated EA-2 families linked to chromosome 19, was also included (*CAN-25*, *-45*, *-191*, *-197*). Fifty randomly collected individuals from the Dutch population (Smith et al., 1988) were used as a control to determine the allele frequencies of polymorphic sites.

Patients with migraine with or without aura were diagnosed according to the international Headache Society (IHS) classification criteria. Patients attending the neurology outpatient clinic of Leiden University Medical Center, The Netherlands, and patients responding to calls in local newspapers or in the periodical of the Dutch Migraine Patients Association, were screened for a positive family history of migraine. Only families with migraine in at least two generations were asked to participate. Probands (*n*=36) and relatives (*n*=492) were personally examined and interviewed using semi-structured questionnaires. The questionnaire included questions about age at onset, frequency and duration of attacks, aura symptoms, premonitory signs and symptoms, triggers for attacks, medication, and additional headaches. When family members were not available for a personal interview, information on their migraine was collected by interviewing their relatives. Because of the low

validity of diagnosing migraine auras through relatives, we only assessed the presence or absence of migraine headaches. Whenever possible, medical records were examined.

5 Genomic structure

Ten different cosmids from the contig extending the invented gene, were subcloned separately in either M13 or pBlueScript KS vector. From each cosmid library at least 3x96 random clones with an average insert size of about 2 kb, were
 10 picked. Positive clones were identified by hybridization techniques and subsequently sequenced with vector-specific primers; intron-exon boundary sequences were completed using cDNA-based primers.

15 Mutation analysis, DHPLC and SSCP

Genomic DNA was used as template to generate polymerase chain reaction (PCR) products for single-strand conformational polymorphism (SSCP) analysis and denaturing high-performance liquid chromatography (DHPLC).
 20 Amplifications were performed in standard conditions with primer pairs as listed in Table 1 or listed below. Except for the 5' side of exon 6, primers were chosen to produce fragments that contained a single exon and at least 35 basepairs (including primer) of each flanking intron
 25 sequence. Amplification of exons 1 and 20 was performed producing two overlapping fragments and exon 19 was amplified into three overlapping fragments. In addition, the following markers;

30	D10S191	Primer sequence 1	CTT TAA TTG CCC TGT CTT C
		Primer sequence 2	TTA ATT CGA CCA CTT CCC
	D10S245	Primer sequence 1	AGT GAG ACT CGT CTC TAA TG
		Primer sequence 2	ACC TAC CTG AAT TCC TGA CC
35	D10S89	Primer sequence 1	AAC ACT AGT GAC ATT ATT TTC A
		Primer sequence 2	AGC TAG GCC TGA AGG CTT CT

DHPLC (Oefner et al., 1995; Hayward et al., 1996) was carried out on automated HPLC instrumentation. Crude PCR products, which had been subjected to an additional 3-minute 95°C denaturing step followed by gradual reannealing from 5 95-65°C over a period of 30 minutes prior to analysis, were eluted with a linear acetonitrile (9017-03, J.T. Baker, Phillipsburg, N.J., USA) gradient of 1.8% per minute at a flow-rate of 0.9 ml/min. The start- and end-points of the gradient were adjusted according to the size of the PCR 10 products (Huber et al., 1995). The temperature required for successful resolution of heteroduplex molecules was determined empirically by injecting one PCR product of each exon at increasing mobile phase temperatures until a significant decrease in retention was observed.

15 For SSCP analysis, primary PCR products were labeled by incorporation of [α -³²P]dCTP in a second round of PCR. Samples were diluted and denatured in formamide buffer before electrophoresis. SSCP was carried out according to published protocols (Orita et al., 1989; Glavac et al., 1994). 20 Digestion of several exons to yield products suitable for SSCP analysis.

Sequencing of PCR products was performed with an ABI 377 automated sequencing apparatus with cycle sequencing according to the manufacturer. Furthermore, PCR products were 25 cloned in the TA vector (Invitrogen) and subjected to manual dideoxy sequence analysis (T7 Sequencing kit, Pharmacia Biotech.).

A total of 481 blood samples were collected from patients with migraine. Genomic DNA was isolated as described 30 by Miller et al., 1988. The analyses of the highly informative microsatellite markers D19S391, D19S394, D19S221 and D19S226, D10S191, D10S245 and D10S89 were performed by PCR; primer sequences related to these markers are available through the human Genome Data Base (GDB).

35 The relative frequencies of marker alleles were estimated on the entire family material, with the relevant

correction for genetic relationships between individuals (Boehnke, M, 1991) with the ILINK option of the I-INKAGE package, version 5.03 (Lathrop et al., 1985). The following marker order and recombination frequencies were used in the

5 multipoint sib-pair analysis: D19S391-5%-D19S394-3%-D19S221-5%-D19S226. Affected-sib-pair analysis was performed using the MAPMAKER/SIBS software package, simultaneously including marker information for all four DNA markers (Kruglyak, 1995). Separate analyses were performed for migraine with aura,

10 migraine without aura, and a combination of both. Allowance was made for dominance variance. When more than two affected sibs occurred in a single sibship, weighted scores were computed according to Suarez and Hodge (1979).

In a sib-pair analysis, the occurrence of parental

15 marker alleles is compared among sibs. Normally, 25% of sib pairs share their marker alleles from both parents, 50% share one marker allele from one of their parents, while the remaining 25% share no parental allele. Deviations from this pattern towards increased sharing, and consistent with the

20 constraints of Holmans's (1993) possible triangle, are explained as linkage (expressed as the maximum lod score MLS). Increased sharing of marker alleles thus indicate that the marker is located closely near a genetic risk factor. The relative-risk ratio for a sib (λ_r), defined as the ratio of

25 the prevalence of a disease in sibs of affected individuals, divided by the prevalence of a disease in the population, can be calculated (May et al., 1995). In other words:

$$\lambda_p = \frac{\text{Affected risk for sib of a proband}}{\text{Affection risk for an individual in the general population}}$$

30

Results

35 Genomic structure

The combination of hybridization and PCR strategies resulted in a rapid assembly of the complete coding sequence

of the human cDNA, with an open reading frame of 6783 nucleotides encoding 2261 amino acid residues (figure 4). The spatial distribution of the human Ca^{2+} channel expression was assayed in rhesus monkey tissues. Total RNA was isolated from several tissues, including various brain structures, and probed with a human cDNA fragment. The probe detected a major transcript of approximately 9.8 kb in cerebellum, cerebral cortex, thalamus and hypothalamus, whereas no transcript was detected in heart, kidney, liver or muscle. There was also no hybridization signal found in RNA preparations from mouse skin tissue or from human peripheral lymphocytes. In addition, an attempt to amplify parts of the cDNA from human peripheral lymphocytes failed.

Complete alignment between the cDNA and individual exon sequences was achieved, allowing the establishment of the exon-intron structure (Table 1). The reconstruction of the exon-intron structure of the CACNL1A4 gene revealed 47 exons ranging in size from 36 bp (exon 44) to 810 bp (exon 19). The exons are distributed over about 300 kb at genomic DNA level. The result shows that the first 10 exons are located in a region of about 150 kb covered by the first 5 cosmids of the contig indicating relatively large introns at 5' side of the gene. Sequences (Figure 1) were obtained of all exons including approximately 100 bp of flanking introns, except for intron 5 adjacent to exon 6. The forward primer of exon 6 harbours the splice junction and 3 bp of exon 6. Splice sites around all exons are compatible with consensus sequence with the exception of splice donor and acceptor of the first intron.

The cosmid contig that yielded the initial Ca^{2+} channel gene exons was extended to cover more than 300 kb. Hybridization experiments showed that the first and last cosmids of the contig were positive for 3'- and 5'-end cDNA sequences, respectively, indicating a genomic distribution of the gene over at least 300 kb (Figure 2). The cosmid contig has been placed into the LLNL physical map of chromosome 19 at band p13.1, between the markers D19S221 and D19S226

(Figure 2). We identified a new polymorphic CA-repeat sequence (D19S1150) on the cosmid contig. Oligonucleotide primers (Table 1) flanking the repeat were synthesized and amplification was performed by PCR as described. Analysis of
5 D19S1150 in 45 random individuals from the Dutch population revealed nine alleles with an observed heterozygosity of 0.82. This highly polymorphic marker is located within the gene and is therefore very useful in genetic analysis.

10 Mutation analysis

Exons and flanking intron sequences, containing the complete coding region of CACNL1A4 and part of untranslated sequences, were screened for the presence of mutations by SSCP and DHPLC analysis in 20 individuals with either FHM or
15 EA-2. Several synonymous nucleotide substitutions and polymorphisms were identified including a highly polymorphic (CAG)_n-repeat in the 3' untranslated region of exon 47 (Table 2). Of all polymorphisms only one was identified predicting an amino acid change, an alanine to threonine
20 substitution at codon 454 (A454T).

Four different missense mutations were found in FHM patients of which one mutation was observed in two unrelated FHM affected individuals (Table 3). The mutations were shown to segregate with the disease within the families; and were
25 not present in about 100 control chromosomes. A G-to-A transition was identified in family *Italy-II* at codon 192, resulting in a substitution of arginine to glutamine (R192Q) within the first voltage sensor domain (IS4). A second missense mutation occurs at codon 666, within the P-segment
30 of the second repeat replacing a threonine residue for methionine (T666M) in family *USA-P*. Two other mutations are located in the 6th transmembrane spanning segment of respectively repeat II and IV. The IIS6 mutation is a T-to-C transition at codon 714, resulting in a substitution of
35 valine to alanine (V714A), identified in FHM family *UK-B*. The mutation in domain IVS6 is an A-to-C transversion at codon 1811 resulting in a substitution of isoleucine to leucine

(I1811L). This I1811L mutation is found in family NL-A and family USA-C, two unrelated FHM families. Comparison of haplotypes in this region, including intragenic markers, did not reveal any evidence for a common founder of family NL-A and USA-C (data not shown). No mutation was found in FHM family Italy-I nor in the six sporadic hemiplegic migraine patients. In addition to missense mutations in FHM families, we also identified mutations in two out of four EA-2 families (Table 3). In EA-2 family CAN-191, a basepair deletion occurs in exon 22 at nucleotide position 4073 causing a frameshift and a premature stop. The second EA-2 mutation is a transition of G-to-A of the first nucleotide of intron 24, predicted to leading to an aberrant splicing in family CAN-26. The invented gene also contains a CAG repeat, of which expansions have been found in patients with autosomal dominant cerebellar ataxia (SCA6). Hence FHM, EA-2 and SCA6 are allelic ion channel disorders and different mutations are associated with different clinical symptomatologies.

Our study patients with common migraine (with or without aura) included 36 independent multigenerational Dutch families. At least some data were available on 937 family members and 212 persons who "married-in" (spouses). Of these, 442 family members (247 affected) and 86 spouses (7 affected) were personally interviewed. The distribution of the different types of migraine among the 247 affected family members are as follows: 132 family members showed migraine without aura, 93 showed migraine with aura and 22 showed months-migraine, not fulfilling all criteria by IHS. Among the 7 affected spouses these figures were 4, 1 and 2, respectively. We scored the parental transmission of migraine in the 36 families (Table 4) to investigate if an additional X-linked dominant or mitochondrial gene was involved. An approximately 2.5:1 preponderance of females among the migraine sufferers was noted, which remained in the affected offspring. Affected fathers were found to transmit migraine

to their sons in 21 cases, making X-linked dominant or mitochondrial inheritance highly unlikely.

The genetic analysis included 204 potentially affected sib pairs; after correction for more than one sib pair in a single sibship the total number of sib pairs was 108. Affected-sib-pair analysis was performed for sib pairs who were both affected with any form of migraine and, in separate analyses, for sib pairs who were both suffering from either migraine with aura or migraine without aura. The informativeness of the region between the markers D19S391, D19S394, D19S221 and D19S226 varied between 82% and 96%. The combined analysis of migraine with and without aura resulted in a maximum multipoint lod score of 1.69 ($p \approx 0.005$) with marker D19S226. For migraine with aura the maximum multipoint lod score was 1.29 corresponding with $p \approx 0.013$ with marker D19S394. The maximum lod score for migraine without aura was not significant (MLS < 0.25) (data not shown). The relative risk ratio for a sib to suffer from migraine with aura (λ_R), defined as the increase in risk of the trait attributable to the 19p13 locus, varied between $\lambda_R = 1.5$ (for marker D19S394) and $\lambda_R = 2.4$ (for marker D19S226). When combining migraine with and without aura, λ_R was 1.25. In a selected portion of 36 Dutch families with migraine with aura and without aura, affected sib-pair analysis was performed for sib pairs who were affected with any form of migraine. The following markers, flanking the $\beta 2$ (CACNB2) calcium channel subunit gene on chromosome 10p12, were tested: D10S191, D10S246 and D10S89. For the combined phenotype (migraine with and without aura) a maximum multipoint lod score of 0.95 ($p < 0.01$) was obtained with marker; D10S191. This result gives independent evidence for a role of the P/Q type Ca^{2+} channel in migraine and other neurological disorders.

Discussion

The genomic structure of the exemplified invented gene revealed 47 exons distributed over about 300 kb (Table 1; Figure 1). A comparison of the gene structure to already

known Ca^{2+} channel $\alpha 1$ subunit genes (CACNL1A1, CACNL1A2, and CACNL1A3) (Soldatov, 1994; Yamada et al., 1995; Hogan et al., 1995), reveals a similar number of exons (50, 49, and 44 respectively) but a larger genomic span (300 kb vs
5 90-150 kb). Remarkably, all splice sites are according to consensus sequence except for intron 1. Splice donor as well as splice acceptor of the first intron do not contain the expected gt...ag intron sequence. An incorrect cDNA sequence is unlikely because the cDNA sequence containing the junction
10 of the first two exons is identical to rabbit and rat sequence. Sequences corresponding to splice donor and acceptor are present in exon 1 and 2, suggesting an additional (yet unidentified) exon in the first intron encompassing part of sequences of exon 1 and exon 2.

15 To test the possible involvement of the invented gene relating to the Ca^{2+} -channel sub-unit in migraine FHM, SCA6 and EA-2, we performed a mutation analysis by DHPLC and SSCP and found several alterations (For example Table 2 & 3). Only one missense variation was observed also present in 2% of the
20 normal controls (Table 2). This polymorphism is a alanine to threonine substitution at codon 454 (A454T), located in the intracellular loop between IS6 and IIS1 (Figure 2). This region contains a conserved alpha interaction domain (AID) that binds subunits (De Waard et al., 1996). However, A454T
25 is located outside the AID consensus sequence and is not likely to be involved.

The identification of two mutations that disrupt the predicted translation product of the invented gene in two unrelated EA-2 patients and the segregation of these
30 mutations with the episodic ataxia phenotype in their families provide strong evidence that the invented gene is the EA-2 gene. A basepair deletion leads to a frame-shift in the putative translation product and encounters a stop codon in the next exon. The frame-shift in this EA-2 family is
35 predicted to yield a calcium channel $\alpha 1$ subunit polypeptide consisting of repeat I and II, and a small portion of repeat III (IIIS1). The G-to-A transition of the first nucleotide of

intron 24 is affecting the nearly invariant GT dinucleotide of the intronic 5' splice junction. The brain-specific expression of the exemplified invented gene makes it extremely difficult to test the hypothesis that this mutation produces aberrantly spliced RNAs by retaining the intron or utilizing other cryptic 5' splice sites.

The frameshift and splice site mutations in EA-2 may suggest a dominant negative effect of the truncated proteins by overruling the (corresponding) intact $\alpha 1$ subunits.

No mutations were found in the remaining EA-2 families (CAN-25 and -197). The use of two independent techniques for mutation screening (DHPLC and SSCP) makes it unlikely that we missed a heterozygote PCR product. Mutations in the promoter region or in intron sequences, resulting in aberrant splicing, may have been the cause of EA-2 in these families. We could also have missed a mutation around the splice acceptor site of intron 5, covered by the forward primer of exon 6. However, larger deletions of e.g. complete exons with flanking intron sequence will disturb the predicted translation product, like the ΔC_{4073} and splice site mutation do, but this is not detectable by a PCR-based screening method but can be seen Southern blot analysis instead.

Four different missense mutations were identified in five unrelated FHM families. These mutations all segregate with FHM within a family and are not observed in over 100 normal chromosomes. The first missense mutation that we describe in the exemplified invented gene occurs in the IS4 domain of the $\alpha 1$ subunit (Table 3; Figure 3). The S4 domains are postulated to be voltage sensors because they have an unusual pattern of positively charged residues at every third or fourth position separated by hydrophobic residues (Tanabe et al., 1987). In calcium channels the positively charged amino acid is an arginine residue (Stea et al., 1995). The mutation in FHM family *Italy-II* predicts a substitution of the first arginine in the IS4 segment with a neutral, non-polar glutamine (R192Q). The change of the net positive

charge of this conserved region of the protein may influence correct functioning of the voltage sensor.

The second missense mutation in FHM family *USA-P* occurs in the P-segment of the second transmembrane repeat. A C-to-T transition predicts substitution of a threonine residue with methionine at codon 666 (T666M). Various observations have shown that P-segments, the hairpin between S5 and S6 that spans only the outer part of the transmembrane region, form the ion-selectivity filter of the pore and binding sites for toxins (Guy and Durell (1996)). Alignment of protein sequence of different P-segments indicating that some residues occur in many different channel genes (Guy and Durell, 1996). The T666M substitution alters one of the conserved residues in the P-segment. It is conceivable that an alteration of a P-segment affects the ion-selectivity or toxin binding of a channel gene.

The remaining two missense mutations identified in FHM families alter the S6 segment of the second and the fourth repeat. A valine to alanine substitution in FHM family *UK-B* is found in domain IIS6 at codon 714 (V714A). Domain IVS6 is mutated in two unrelated FHM families (*NL-A* and *USA-C*), predicting a isoleucine to leucine substitution at codon 1811 (I1811L). The V714A and I1811L missense mutations do not really change the neutral-polar nature of the amino acid residues. However, both S6 mutations are located nearly at the same residue at the intracellular site of the segment and are conserved in all calcium channel $\alpha 1$ subunit genes. In addition, the A-to-C transversion leading the I1811L substitution occurred in two unrelated FHM families on different haplotypes indicating recurrent mutations rather than a founder effect. Although the exact function of the S6 domains are not known, these data strongly suggest that mutations in IIS6 and IVS6 result in FHM.

The I1811L mutation is present in two FHM families of which one (*NL-A*) also displays a cerebellar atrophy in (some) affected family members. The presence of cerebellar atrophy in FHM families has been reported in about 40% of chromosome

19-linked FHM families, whereas none of the unlinked families was found to have cerebellar atrophy (Terwindt et al., 1996).

The I1811L mutation excludes the possibility of allelic mutations in FHM and FHM with cerebellar atrophy. However, it is likely that FHM or FHM with cerebellar atrophy are the result of pleiotropic expression of a single defective gene.

No mutation was found in a small Italian FHM family (*Italy-I*). Apart from the possibilities discussed for EA-2, it should be noted that linkage to 19p13 was only suggested but never proved with significant lod scores (M. Ferrari, personal knowledge).

The four missense mutations identified indicate a mechanism for FHM in which both alleles of the α_1 subunit are expressed, one harbouring an amino acid substitution which affects the function of this calcium channel α_1 subunit by reducing or enhancing the electrical excitability. The relationship of FHM and other types of migraine makes it highly rewarding to investigate the involvement of the only missense variant observed (A454T) (Table 2), and to continue the search for other variants of the exemplified invented gene specific for common types of migraine.

The mutations in EA-2 and FHM demonstrate among others that the brain specific calcium channel gene CACNL1A4 is responsible for both EA-2 and FHM, and is also involved in the primary pathogenesis of the more common forms of migraine. We conducted the common migraine study in an independent sample of 36 extended Dutch families, with migraine with aura and migraine without aura. We found significant increased sharing of the marker alleles in sibs with migraine with aura (MLS=1.29 corresponding with $p \approx 0.013$). Although no such increased sharing was found for migraine without aura, a combined analysis for both migraine types resulted in an even more significant increased sharing (MLS=1.69 corresponding with $p \approx 0.005$). These results clearly indicate the involvement of the calcium α_{1A} -subunit gene region on 19p13 in both migraine with and without aura; the contribution to migraine with aura, however, seems strongest.

The positive findings in our study clearly demonstrate an involvement of the FHM locus region in non-hemiplegic familial migraine, notably in migraine with aura. The P/Q-type calcium channel α_{1A} -subunit gene on chromosome 5 19p13 may be an "aura-gene" and is involved in both FHM and migraine with aura, but not in migraine without aura. This however, seems unlikely since an increased sharing of marker alleles was also found when we combined the results for migraine with and without aura. Furthermore, the increase in 10 sharing was stronger than expected to be only due to the contribution of migraine with aura. An alternative explanation is that the gene is involved in both types of migraine, but in migraine without aura there is an additional strong effect of other, possibly environmental factors, 15 thereby reducing the penetrance.

The latter view is also supported by the results obtained from calculating the relative risk ratios (λ_r) for sibs from affected individuals to also have migraine. The relative risk ratio for a sib to suffer from migraine with 20 aura was $\lambda_r=2.4$. When combining migraine with and without aura, λ_r was 1.25. In a population-based study the relative risk for first degree relatives of probands with migraine with aura to also have migraine with aura was $\lambda_r=3.8$. Because of the female preponderance among migraine patients, X-linked 25 dominant or mitochondrial inheritance has been suggested to be involved in familial migraine. Although a predominant maternal inheritance pattern was noted in our families, X-linked dominant or mitochondrial inheritance were found to be highly unlikely because affected fathers transmit migraine to 30 their sons. Furthermore, the predominant maternal inheritance can be explained by the female preponderance among the migraine patients.

We conclude that the well-established genetic contribution to the etiology of migraine is partly, but not 35 entirely, due to genetic factors located in the chromosomal region of the P/Q-type calcium channel α_{1A} -subunit gene. Further analysis of the cerebral distribution and function of

this calcium channel, as well as of the "mutated channels", will help to unravel the pathogenetic pathway of migraine. It may also contribute to a better understanding of the mechanisms involved in related disorders such as episodic ataxia type-2, autosomal dominant cerebellar ataxia (SCA6), cerebral atrophy, and epilepsy, which all have been found to be associated with mutations in this gene. Study of FHM, EA-2 mutants and variants such as the A454T variant expressed in vitro or in mouse or other experimental animal models will ultimately lead to better understanding of the diseases, their cellular mechanisms, and the clinical relationship between FHM, EA-2, migraine, and other episodic neurological disorders such as epilepsy, and will provide rationales for the development of prophylactic therapy.

15 **Localization and identification of the mouse gene related to the neurological mouse mutations tottering, leaning and rolling.**

The tottering (tg) mutation arose spontaneously in the DBA inbred strain, and has been back-crossed into a C57BL/6J (B6) inbred strain for at least 30 generations. The genome of the tg mouse therefore is of B6 origin except for a small region around the tg gene on chromosome 8. Interestingly, the chromosome 8 region in mouse has synteny with the human chromosome 19p13.1, in which the human calcium channel alpha1 subunit has been identified. We therefore consider the tg locus as a possible site of the mouse homologue of the human calcium channel gene.

To determine the exact localization of the mouse homologue, PCR was carried out with primers based on human cDNA sequence selected from Figure 1 and mouse genomic DNA as template. In human, primers were known to be located in different flanking exons. PCR amplification on human DNA yielded a 1.5kb fragment.

35 Forward primer : 5'- caa cat cat gct ttc ctg cc-3'

Reversed primer: 5'- atg atg acg gcg aca aag ag-3'

Amplification on mouse DNA yielded a 750-bp fragment. The fragment mainly consists of intronic sequences. SSCP analysis revealed several polymorphisms in the different inbred strains (each strain a specific pattern). Analysis of
5 amplified product of the tg/tg (homozygote) and tg/+ (heterozygote) mice demonstrated a DBA specific signal in the tg/tg mouse, and a heterozygous pattern of DBA and B6 inbred strains in the heterozygous tg/+ mouse. These results show that the mouse homologue of the human
10 calcium channel $\alpha 1$ subunit is located within the mouse tottering interval on chromosome 8.

In conclusion: the phenotypic characteristics of the tg mouse (tg/tg and tg/+) suggest involvement of ion-channels in the tg-etiology. The localization of the mouse homologue of
15 the human calcium gene within the tottering interval show that a tottering phenotype in mouse is caused by a mutation in the mouse homologue of the CACNL1A4 gene. With various variants of the tottering mouse (the Jackson Laboratory, Bar Harbor, ME, USA), such as the leaner and rolling varieties,
20 such mutations in the mouse homologue of the CACNL1A4 gene can be found, clearly demonstrating that the gene is related to a variety of episodic neurologic disorders and using this genetic information one can engage in a variety of pathophysiological studies, as for example indicated below.

25 The tg mutation arose spontaneously in the DBA/2 inbred strain. tg/tg homozygotes are characterized by a wobbly gait affecting the hindquarters in particular, which begins at about 3 to 4 weeks of age, and by intermittent spontaneous seizures which resemble human epileptic absence seizures. The
30 central nervous system of the mice appears normal by light microscopy. There is no discernible cerebellar hypoplasia. In fluorescent histochemistry studies tg/tg mice show a marked increase in number of noradrenergic fibers in the terminal fields innervated by locus ceruleus axons, the hippocampus, cerebellum, and dorsal lateral geniculate. Treatment of
35 neonatal tg/tg mice with 6-hydroxydopamine, which selectively causes degeneration of distal noradrenergic axons from the

locus ceruleus, almost completely abolishes the ataxic and seizure symptoms.

The leaner mutation of the tottering mouse arose spontaneously in the AKR/J strain. Homozygotes are recognized
5 at 8 to 10 days of age by ataxia, stiffness, and retarded motor activity. Adults are characterized by instability of the trunk, and hypertonia of trunk and limb muscles. The cerebellum is reduced in size, particularly in the anterior region, in tg<la>/tg<la> mice, as is the case with a certain
10 number of FHM patients. There is loss of granule cells beginning at 10 days of age and loss of Purkinje and Golgi cells beginning after 1 month. Cell loss later slows but continues throughout life. Granule and Purkinje cells are more severely affected than Golgi cells and the anterior
15 folia more severely affected than other parts of the cerebellum. The cerebellum of tg<la>/tg mice shows shrinkage and degenerative changes of the Purkinje cells. The loss in cerebellar volume in tg<la>/tg and in tg/tg mice is specific to the molecular layer, with no change in volume of the
20 granule cell layer or the white matter layer. Allelism of leaner with tottering was shown in complementation and linkage tests.

A third variety of the tottering mouse is (tg<rol>) called the rolling Nagoya. Found among descendants of a cross
25 between the SIII and C57BL/6 strains, the tg<rol> mutation apparently occurred in the SIII strain. Homozygotes show poor motor coordination of hindlimbs that may lead to falling and rolling, and sometimes show stiffness of the hindlimbs and tail. No seizures have been observed. Symptoms are
30 recognizable at 10 to 14 days old. They appear a little earlier than those of tg/tg mice and are somewhat more severe. The cerebellum is grossly normal until 10 days of age, but after that grows more slowly than normal. The size of the anterior part of the central lobe of the cerebellum is
35 reduced with reduction in the numbers of granule, basket, and stellate cells but normal numbers of Purkinje cells. There is a reduced concentration of glutamate and an increased

concentration of glycine and taurine in the cerebellum and decreased activity of tyrosine hydroxylase in the cerebellum and other areas.

Legends to figures

Figure 1

5 Nucleic acid sequences of 47 exons and flanking intron
sequences containing the complete coding region of the
invented gene and part of untranslated sequences.

Figure 2

10 Genetic map, cosmid contig and global exon distribution of
the invented gene on chromosome 19p13.1. The cosmid contig is
shown with EcoRI restriction sites, available via Lawrence
Livermore National Laboratory; exon positions are indicated
schematically, regardless of exon or intron sizes (Table 1).
D19S1150 is a highly polymorphic intragenic (Ca)_n-repeat.

15

Figure 3

Membrane topology of $\alpha 1$ subunit of the P/Q-type Ca^{2+} -channel.
The location and amino acid substitutions are indicated for
mutations that cause FHM or EA-2

20

Figure 4

The coding sequence of human cDNA of the invented gene with
an open reading frame encoding 2261 amino acid residues.

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Exon/intron organization of the human invented gene
and exon-specific primer pairs

Table 1

Exon	cDNA	Size	Domain	Cosmid(s)	Primer Forward	Primer Reversed	Size
1	UTR - 568	500		25960 / 30151	tct ccg cag tgg tag ctc ca cgc aaa gga tgt aca agc ag	ggg tgt aga gtc cca tgg tc att ccc aag cct cca ggg tag	320 370
2	569 - 674	106	I S1	30151	cac ctc caa cac cct tct tt	tct gtc ccc tgc tcc act c	240
3	675 - 814	140	I S2, I S3	30151	acg ctg acc ttg cct tct ct	caa cca aaa gcc tgg taa tc	230
4	815 - 906	92	I S3, I S4	28913	aaa acc cac cct ctg ttc tc	ttg tca ggg tgg gaa act ca	160
5	907 - 1059	153	I S5	28913 / 27415	ctt ggt ggc ggg gtt t	ctg cct aat cct ccc aag ag	290
6	1060 - 1253	194		27415	tcc ctt ccc tt tgt aga tg	gtg ggg ctg tgt tgt cct t	350
7	1254 - 1357	104	I S6	27415	gac aga gcc aca aga gaa cc	agc aaa gag gag tga gtc gg	250
8	1358 - 1473	116		34077 / 27415	ata ctc tgg ctt ttc tat gc	gca tga ctc tct ttg tac tc	230
9	1474 - 1530	57		34077	gca gag aat ggg ggt gg	ctg agg tgg gtt tag agc ag	180
10	1531 - 1623	93		34077	ggg taa cgt ctt tt ctc ttg c	atg tct ctt ggg cga tag gt	200
11	1624 - 1833	210	II S1	16894 / 32236	att tct tct gaa gga aca gc	gga ggg atc agg gag ttg gc	310
12	1834 - 1946	113	II S2, II S3	16894	caa gcc taa cct cct ctc tg	tca ttc cag gca aga gct g	200
13	1947 - 2051	105	II S3, II S4	16894	att tgg agg gag gag tt gg	tca ctt tcc caa ctt tct gg	310
14	2052 - 2191	140	II S4, II S5	16894	cag aaa gtt ggg aaa gta gc	ttg aat tcc tgt gaa gga c	250
15	2192 - 2264	73		16894	ctt gga gat gag ata ctg agc	cag gca ctt tca tct gtc ac	200
16	2265 - 2382	118	II S6	16894	tcc aca gct gca tct cca ag	acc ctc cct tga gcc cct	270
17	2383 - 2450	68	II S6	16894	cag tgg ttg ctt ttc ctg ac	ttg cca gag aaa cat tct cc	130
18	2451 - 2557	107		16894	tga aca aag att cca cgt gc	ttc agg agc cag ggt agc atc	170
19	2558 - 3367	810		16894	tag caa tgc tct aag tcc tg cgc agg aga acc gca aca a gc agc agg gag agc cgc agc	tgt ttc ctg agg aag tcc tc gcg atg acg tgg atg ctc tac cgt cat tct gcg gat tc	320 450 300
20	3368 - 3831	464		16894	ggg tct tt tca ttc act tgc gag aat agc ctt atc gtc ac	tt cct ggc agt ctt agc tg cag tga tgt gag agc aga g	430 200
21	3832 - 3973	142		16894 / 34275	tgg gaa att gtc gag gga gc	tga ctt ccg cca ccc tgg tg	250
22	3974 - 4103	130	III S1	16894 / 34275	agc ctg tgg tct gag tgg ac	tag gaa ggg gtc tgc tct gtc	210
23	4104 - 4163	60	III S2, III S3	16894 / 34275	atc cac tgc tct ctt gct tt	gtg gtt ctc act tat aat ctg c	170
24	4164 - 4270	107	III S3	34275	tgg cct cat tgg ctt ccc tgc	aag agg aaa ccc ttg cga ag	250
25	4271 - 4370	100	III S4	34275	cta ccc aac ctg acc tct gc	aca tga taa ccc tga cag tc	220
26	4371 - 4531	161	III S5	34275	ctc atg ctc tct gtc aac tc	tgg ttc caa tgg gaa tgt gc	250
27	4532 - 4669	138		34275	ctg ctt ccc aag cag tct ag	tcc tgg ata gat ttc cag tc	300
28	4670 - 4871	202	III S6	34275	agt tt taa agg aca gat gg	tt ccc tgc ccc att cct ttg c	280
29	4872 - 5036	165	IV S1	34275	ctc tgc cgc tct cac cac tg	tt atc agg tag agg cag g	250
30	5037 - 5147	111	IV S1, IV S2	34275	ttc caa gcc cat agc tgt agc	tga ccc tgc tac tcc tgc ttc	180

31	5148 - 5231	84	IV S3	15496	act gtg cct cta aca tgc ac	aag tgc tgg ctg aag cag	250
32	5232 - 5348	117	IV S4	15496	tct gtg agt ggt gac agc tc	gtc acc tgt ctt ctg agc	240
33	5349 - 5414	66	IV S5	15496	tgg aag gac tct ggc acg tg	gga ggc tct ggg aac ctt ag	250
34	5415 - 5530	116		15496	aga agc cac tgg agg aat ggc	att atc aga gca ggt ccc ctt c	250
35	5531 - 5681	151	IV S6	15496	tcc gag tct ctg att tct cc	aga cgg ccc tca cag tgt c	210
36	5682 - 5809	128	IV S6	15496	ttc att ccc tgg gtc tct gc	ctg act gaa cct gtg aga c	350
37	5810 - 5906	97		15496	tgt gaa ccc att gcc tgc a	tgg gaa tga ctg cgc ttg c	200
38	5907 - 6012	106		15496	atg cct ggg aat gac tgc	tgt cac gcc tgt ctg tgc	200
39	6013 - 6120	108		15496	tga cac cca ggc agg cag	tct gtc ctg gtg gat tgg atc	200
40	6121 - 6221	101		15496	tgg gtg agc tca ccg tgt	ttc ccg tgg tga cat gca agc	200
41	6222 - 6331	110		15496	gtc cac aca ctg ctg tct gc	aca ctg cac ctg cct ggc	320
42	6332 - 6470	139		15496	gcc agg gag gtg gag tgt	ggg tcc ttc cac cgc aac	550
43	6471 - 6584	114		15496 / 30762	caa ctg ccc aat ggc tc	cct acc cag tgc aga gtg agg	350
44	6585 - 6620	36		15496 / 30762	tct gtg tgc acc atc cca tg	aag gat tgg gct cca tgg ag	200
45	6621 - 6807	187		15496 / 30762	gtt ggt gct agc tgc tga c	ctt tct tct tcc tta gtg tc	330
46	6808 - 7061	254		15496 / 30762	gtg tgc tgt ctg acc ctg ac	agc ctg ggg tca ctt gca gc	320
47	7062 - UTR	2350		/ 30762	cct ttg ttg caa ttg tgg tgt ag	tgg ggc ctg ggt acc tcc aa	280

Note. Sizes of exons and PCR products are given in basepairs;
domains of protein are indicated according to Stea et al., 1995.

Table 2 Polymorphisms in coding sequence of the invented gene

Location	Nucleotide change			Frequency	Consequence
exon 4	nt 854	G - A	Thr ₁₉₃	0.02	-
exon 6	nt 1151	A - G	Glu ₂₉₂	0.07	-
exon 8	nt 1457	G - A	Glu ₃₉₄	0.38	-
exon 11	nt 1635	G - A	Ala ₄₅₄	0.02	Ala ₄₅₄ - Thr (A454T)
exon 16	nt 2369	G - A	Thr ₆₉₈	0.12	-
exon 19	nt 3029	G - A	Glu ₉₁₈	0.07	-
exon 23	nt 4142	T - C	Phe ₁₂₈₅	0.22	-
exon 46	nt 6938	T - C	His ₂₂₂₁	0.46	-
exon 47	nt 7213	(CAG) _n	3'UTR	#	-

Note. Frequency as observed in 100 control chromosomes: # Seven alleles of (CAG)_n were observed in the range between n=4 to n=14, with a heterozygosity value of 0.75.

Table 3 Mutations of the invented gene in families with FHM or EA-2

<i>Disease</i>	<i>Family</i>	<i>Location</i>	<i>Domain</i>	<i>Nucleotide change</i>		<i>Consequence</i>	
FHM	It-II	exon 4	I S4	nt 850	G - A	Arg ₁₉₂ - Gln (gain of SfcI site)	R192Q
FHM	US-P	exon 16	P-segment	nt 2272	C - T	Thr ₆₆₆ - Met	T666M
FHM	UK-B	exon 17	II S6	nt 2416	T - C	Val ₇₁₄ - Ala (gain of BbvI site)	V714A
FHM	NL-A/US-C	exon 36	IV S6	nt 5706	A - C	Ile ₈₁ - Leu (gain of MnlI site)	I1811L
EA-2	CAN-191	exon 22	III S1	nt 4073	deletion C	frameshift (loss of NlaIV site)	STOP ₂₉₄
EA-2	CAN-26	intron 24	splice site	nt 4270-1	G - A	AC/gt - AC/at (loss of BsaAI site)	aberrant splicing

Table 4. Parental transmission of migraine for 36 unrelated Dutch families.

<i>parents</i>	<i>N</i>	<i>offspring</i>	<i>N</i>	<i>affected</i> <i>N(%)</i>	<i>ratio*</i>
healthy father x migraine mother	51	daughters	72	48 (66.7%)	2.3:1
		sons	72	21 (29.2%)	
migraine father x healthy mother	18	daughters	26	17 (65.4%)	2.5:1
		sons	15	4 (26.7%)	

* ratio of proportion affected sons/proportion affected daughters

CLAIMS

1. An isolated and/or recombinant nucleic acid encoding a Ca^{2+} -channel $\alpha 1$ subunit related to (familial hemiplegic) migraine and/or episodic ataxia type-2 derived from, related to or associated with a gene which in humans is present on
5 chromosome 19p13.1-19p13.2 or a specific fragment or homolog or derivative thereof.
2. A nucleic acid according to claim 1 which is a cDNA molecule.
3. A cDNA molecule according to claim 2 comprising a
10 6800 bp coding region.
4. A nucleic acid according to claim 1, 2 or 3 which is of human origin.
5. A nucleic acid according to claim 4 and showing at least 70% homology with the nucleic acid sequence as listed in
15 figure 1.
6. A nucleic acid according to any of claims 1-5 and showing at least 90% homology with the nucleic acid sequence as listed in figure 1.
7. A nucleic acid according to any of claims 1-6 and
20 showing a mutation at codon 192 resulting in the replacement of arginine by glutamine.
8. A nucleic acid according to any of claims 1-7 and showing a mutation at codon 666 resulting in the replacement of threonine by methionine.
- 25 9. A nucleic acid according to any of claims 1-8 and showing a mutation at codon 714 resulting in the replacement of valine by alanine.
10. A nucleic acid according to any of claims 1-9 and showing a mutation at codon 1811 resulting in the replacement
30 of isoleucine by leucine.
11. A nucleic acid according to any of claims 1-10 and comprising a CA-repeat sequence as shown in figure 2.

12. A nucleic acid according to any of claims 1-11 and comprising a (CAG)_n repeat sequence as shown in figure 2.
13. A nucleic acid according to any of claims 1-12 and comprising a polymorphism in the coding sequence.
- 5 14. A nucleic acid according to claim 13 and comprising a polymorphism in the coding sequence as shown in table 2.
15. A nucleic acid according to claim 13 or 14 and comprising a mutation at codon 454 resulting in a replacement of alanine by threonine.
- 10 16. A nucleic acid according to any of claims 1-15 and comprising a deletion.
17. A nucleic acid according to any of claims 1-16 and comprising a frameshift at codon 1266.
18. A nucleic acid according to any of claims 1-17 and comprising a mutation resulting in aberrant splicing.
- 15 19. A nucleic acid according to any of claims 1-18 and comprising a mutation resulting in aberrant splicing of intron 28.
20. An isolated and/or recombinant nucleic acid encoding a CA²⁺ channel subunit or a functional fragment thereof related to (familial hemiplegic) migraine and/or episodic ataxia type 2.
21. An isolated and/or recombinant nucleic acid encoding a CA²⁺ channel β 2 subunit related to (familial hemiplegic) migraine and/or episodic ataxia type 2, derived from, related to or associated with a gene which in humans is present on chromosome 10p12 or a specific fragment thereof.
- 25 22. A method for localising or identifying a gene using a nucleic acid molecule or a fragment of fragments thereof according to any of claims 1-21.
- 30 23. A method according to claim 22 wherein the gene is related to episodic neurological disorders.
24. A method according to claim 22 or 23 wherein the gene is related to migraine.
- 35 25. A method according to claim 22, 23 or 24 wherein the gene is related to FHM and/or EA-2 and/or autosomal dominant cerebellar ataxia.

26. A method distinguishing between alleles of a gene using a nucleic acid molecule or a fragment of fragments thereof according to any of claims 1-21.
27. A method according to any of claims 23-26 in which the
5 gene is of human origin.
28. A method according to any of claims 23-27 which comprises selecting a cell or an animal.
29. A recombinant expression vector comprising a nucleic acid molecule or a fragment of fragments thereof according to
10 any of claims 1-21.
30. A cell or an animal comprising a vector according to claim 29.
31. A cell or an animal comprising a nucleic acid molecule or a fragment of fragments thereof according to any of claims
15 1-21.
32. A cell or an animal selected by a method according to claim 28.
33. A cell or an animal comprising a genome in which nucleic acid sequences corresponding to nucleic acid molecules
20 according to any of claims 1-21 have been modified.
34. Use of a cell or an animal according to any of claim 30-33 to test or develop specific medication for the treatment of FHM, EA-2, SCA6, migraine or other neurological disorders associated with cation channel dysfunction.
- 25 35. A protein or peptide comprising an amino acid sequence encoded by a nucleic acid molecule, or a fragment or fragments thereof, according to any of claims 1-21.
36. A natural or synthetic antibody directed against a protein or peptide according to claim 35.
- 30 37. Use of a protein or peptide or antibody according to claim 35 or 36 to detect or diagnose FHM, EA-2, SCA6, migraine or other neurological disorders associated with cation channel dysfunction.

Figure 1

```
//
Submission no : 1
exon 1 : <..672
start codon : 381..383
intron 1 : 673.
Remarks : no consensus splice site intron 1
```

tttttttacg	ttctcttttt	tttcgagtg	tgactggatg	ctgattcttc	50
ctcgtatttt	tgctgcttct	ctctccctcc	cctccttccc	gggcccgggc	100
cgcggccgca	ccctccttcc	gccccctcct	ctccgggggc	agccaggaag	150
atgtcccag	ctgctatccc	cggctcggcc	cgggcagccg	ccttctgagc	200
ccccgaccgc	agcgccgagc	cgcgcgcgca	tgggctgggc	cgtggagcgt	250
ctccgcagtc	gtagctccag	ccgcgcgcgt	cccagccccg	gcagcctcag	300
catcagcggc	ggcggcggcg	gcggcggcgt	cttcgcgcgc	gttcgcgcgc	350
gcgtaaccgg	agccctttgc	tctttgcaga	ATGGCCCGCT	TCGGAGACGA	400
GATGCCGGCC	CGCTACGGGG	GAGGAGGCTC	CGGGGCAGCC	GCCGGGGTGG	450
TCGTGGGCAG	CGGAGGCGGG	CGAGGAGCCG	GGGGCAGCCG	GCAGGGCGGG	500
CAGCCCGGGG	CGCAAAGGAT	GTACAAGCAG	TCAATGGCGC	AGAGAGCGCG	550
GACCATGGCA	CTCTACAACC	CCATCCCCGT	CCGACAGAAC	TGCCTCACGG	600
TTAACC GGTC	TCTCTTCCTC	TTCAGCGAAG	ACAACGTGGT	GAGAAAATAC	650
GCCAAAAGAT	CACCGAATGG	CCatatacct	ttgcccgaac	cccagcagca	700
gctgcgcctc	ccccctctcc	ctccgcctcc	cctcttccag	gctgggagag	750
agaccggggg	gttgatggga	ggtggggagg	aggggggtct	tccaggggct	800
gggagagggg	gcaccgggag	gagtgtgaaa	gaatctctcc	accccgagct	850
gggttgagct	accctggagg	cttggggaatg	gggttttcgg	gggctggggg	900
ccggccagcc	ggagagtggg	tccttcccaa	ggaccgactc	tagaatgaga	950
tct					953

//

Submission no : 2
 Intron 1 : <..88
 Exon 2 : 89..194
 Intron 2 : 195..>
 Remarks : No consensus splice site intron 1

gatcttthycc	actgggggtca	gtggggggtgg	gtgcacctcc	aacacccttc	50
ttttctttga	acaagattht	tccttaattc	cccaataacTC	CCTTTGAATA	100
TATGATTTTA	GCCACCATCA	TAGCGAATTG	CATCGTCCTC	GCACTGGAGC	150
AGCATCTGCC	TGATGATGAC	AAGACCCCGA	TGTCTGAACG	GCTGgtgagt	200
gatgtctttt	ctcaggggtct	tctccttggc	tttagcagga	cattaatttt	250
tggggggagt	gagcagggca	cagaggaggc	tctcagtcct	ggagcccaga	300
gccagatcat	gggaagccta	aatttccttt	tcattttttc	ttgaaccaga	350
gtctcgctct	gtcaccacagg	ctggagtgca	gtggttcagt	catagctcac	400
tgcagcctcc	acctcctggg	ctcaagccat	cctcccactg	cagcctcctg	450
agtagcaggg	actaacaggt	gccaccatgc	ccagttaatt	ttcttatttt	500
tatctttttt	tgtaagaaga	tgggggat			527

//
Submission no : 3
Intron 2 : <..57
Exon 3 : 58..197
Intron 3 : 198..>

gatccttg	tca	acatctg	ccc	agcccaag	ac	gctgac	ccttg	ccttctct	ccc	50
cttccag	GAT	GACACAG	AAC	CATACTT	CAT	TGGAAT	TTTT	TGTTTC	GAGG	100
CTGGAAT	TAA	AATCATT	GCC	CTGGGTT	TG	CCTTCC	ACAA	AGGCTC	CCTAC	150
TTGAGGA	ATG	GCTGGA	ATGT	CATGGAC	TTT	GTGGT	GGTGC	TAACGG	Ggta	200
agtggc	gcgt	gctata	cgt	ttggatt	taa	ctagct	gaag	gattac	gagg	250
cttttg	gttg	gtgtgg	tccg	ggccagg	ctc	aggaag	gctg	agccct	tg	300
ttctcc	cctcc	ccttg	tatat	cgcctg	cctc	ctttct	gcca	acacccc	acc	350
tccatg	tctc	agctgt	tatat	tacagc	agat	gctttc	tggt	acaatta	aaaa	400
taatag	ctca	ttattg	ttgg	ctgctt	ccag	agtgc	tttat	g		441

//

Submission no : 4
Intron 3 : <..142
Exon 4 : 143..234
Intron 5 : 235..>

aaaactgagg	ccagtgggtgt	cgagtcacct	gcctgtgggtc	acccaaccaa	50
tacaggacag	cttggaatcc	caagcacccc	cgccctgctg	tctgaccccc	100
aaaaccacc	ctctgttctc	cattctgggt	tctttctttc	agCATCTTGG	150
CGACAGTTGG	GACGGAGTTT	GACCTACGGA	CGCTGAGGGC	AGTTCGAGTG	200
CTGCGGCCGC	TCAAGCTGGT	GTCTGGAATC	CCAAgtgcgt	gagtttccga	250
ccctgacaa					259

//

Submission no : 5
Intron 4 : <..118
Exon 5 : 119..271
Intron 5 : 272..>

cttaatat	cctcagga	acacctgc	tgtctggg	agacctgg	50
gtcttggt	cggggttt	ggggtact	ctcatggg	tatggggc	100
ctctctgt	ccccccag	TTACAAGT	TCCTGAAG	GATCATGA	150
GCGATGAT	CTTTGCTG	GATCGGC	CTCCTATT	TTGCAATC	200
TATTTTTG	ATCATAGG	TAGAATTT	TATGGGAA	TTTCATAC	250
CCTGCTTT	AGAGGGGA	Ggtaggtc	cggagcat	tgcattct	300
cagttttc	cttcaggg	aagctctt	gaggattag	caggggtg	350
cttctttc	ctggcagc	ggaggacc	ctccttcag	gagcacta	399

//
Submission no : 6
Intron 5 : <..22
Exon 6 : 23..216
Intron 6 : 217..>

ttttttccct	tcccttttgt	agATGACATT	CAGGGTGAGT	CTCCGGCTCC	50
ATGTGGGACA	GAAGAGCCCG	CCCGCACCTG	CCCCAATGGG	ACCAAATGTC	100
AGCCCTACTG	GGAAGGGCCC	AACAACGGGA	TCACTCAGTT	CGACAACATC	150
CTGTTTGCAG	TGCTGACTGT	TTTCCAGTGC	ATAACCATGG	AAGGGTGGAC	200
TGATCTCCTC	TACAATgtaa	gtgatgctgg	gacagtgtgt	gtggacaatc	250
agagtctcag	ggaggtggcc	tcctgggacc	agtgagactc	caaggctgca	300
atggagggac	cctgagctgg	gaaaggcagc	ccaaggacaa	cacagcccca	350
ctgaagctgg	cctgaggctc	aggcttttga	agattacagg	ggctcatgag	400
cagaactcta	actatagggc	atagaagtct	ggagggcccc	cagatgcaac	450
atcatttttc	attgtgcaag	tgtttagata	taattttaga	tttttgaata	500
cggaaagggt	atgtgatcca	aaatccaaca	cagataaaaag	atagagtaat	550
atcctttggac	gtaggcgagg	ggtccctgcc	ctgagg		586

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Submission no : 7
Intron 6 : <..183
Exon 7 : 184..287
Intron 7 : 288..>

tttcttcaga	aaacgggttc	ttcctccatt	tccccctctg	ggatgccaga	50
gccccagaac	tccacaagcc	aagaacattt	aagacagagc	cacaagagaa	100
ccgagcttcc	ccttccctca	cctgtcaggt	tctatctgag	tcccagtcaa	150
ctctcacctg	ctttccctcc	tcacacccta	cagAGCAACG	ATGCCTCAGG	200
GAACACTTGG	AACTGGTTGT	ACTTCATCCC	CCTCATCATC	ATCGGCTCCT	250
TTTTTATGCT	GAACCTTGTG	CTGGGTGTGC	TGTCAGGgta	agtttctgct	300
actccccacc	ccatcccact	cactcctctt	tgctaaacttc	tttccaagta	350
gaggccattg	aagctttggt	ttcattcact	agacaga		387

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//
Submission no :      8
Intron 7      :      <..190
Exon 8        :      191..306
Intron 8      :      307..>
Sequence      :      412
Remark        :      intron 7 contains CA-repeat (D19S1150)
```

cccagtccttt	tcccagaagt	cctgactcct	cctggttgaaa	actcctgacc	50
tccagggact	tctgaatccc	caaacacaca	cacacacaaa	cacacacaca	100
cacacacaca	cacacacaca	caaacacaca	cacaaacgtt	tcctaacatt	150
tt-aaaacag	ccatactctg	gcttttctat	gcttctccag	GGAGTTTGCC	200
AAAGAAAGGG	AACGGGTGGA	GAACCGGCGG	GCTTTTCTGA	AGCTGAGGCG	250
GCAACAACAG	ATTGAACGTG	AGCTCAATGG	GTACATGGAA	TGGATCTCAA	300
AAGCAGgtga	ggccctttca	tcctggggcc	cagggatgga	gatcccaggc	350
cacagagtac	aaagagagtc	atgcagtttg	gagaaggcta	agctgggagg	400
gttatgatgg	ga				412

//

Submission no : 9
 Intron 8 : <..513
 Exon 9 : 514..570
 Intron 9 : 571..>

gagtaggaag	ttagaggcag	ggtggtcagg	gaaggcttct	ctaaggaagt	50
accctctgag	cagagagacc	tgaaggacgt	gaagaaggaa	gctgtgggga	100
tgtcaagga	aggggcatc	caggcagaga	cagcaagtgc	aaaggccctg	150
agctaggaac	gtatgtgaga	cacagcaagg	aagccagtgc	agctgaaaca	200
gagtgaagag	tggggacagc	tggaggagag	gaagacagga	aggtgatgga	250
gatcagatca	agcaggggct	tataggctgt	ggtgtggaca	ttggttttta	300
ttttgcgcga	ggtggggaga	atgttggcta	ttgctactgt	tgcgagggtg	350
gggcttgaag	tcacaaacca	cccagcagca	tgttttttgg	tcggttgagc	400
tgtcaccatc	agtcagcaga	gaatgggggt	ggccgggcag	acccttcttc	450
ctgggtccaag	ggagaactca	tcctccaaat	gcaggagctt	aactctgtgc	500
tcttctctt	cagAAGAGGT	GATCCTCGCC	GAGGATGAAA	CTGACGGGGA	550
GCAGAGGCAT	CCCTTTGATG	gtaactgctc	taaaccacc	tcaggggtgg	600
gtcccagggg	a				611

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Submission no : 10
Intron 9 : <..86
Exon 10 : 87..179
Intron 10 : 179..>

ttaatccaag	acacactgtg	tgctctatat	ggctctgtgtt	cgaaaaaggg	50
taacgtcttt	ttctcttgcc	atgtttccat	tgtagGAGC	TCTGCGGAGA	100
ACCACCATAA	AGAAAAGCAA	GACAGATTTG	CTCAACCCCG	AAAGAGGCTGA	150
GGATCAGCTG	GCTGATATAG	CCTCTGTGGg	tgagtccctt	cctctgccac	200
ctatcagttg	ttcatcacct	atcgcccaag	agacatgggtg	gggtgggggc	250
agagggcttg	caaaccgtgc	tgcttggtt	tgggtctcag	ctccaccctt	300
tcccacctgt	gcgtgtgtcc	tgggcagatt	acatcattat	gggaataaca	350
tccgtgccta	gcttctcatt	atgttggtgg	aattcaacta	aatgatcccc	400
atgaagcatg	gcaaaccagc	acctggcagg	gacgaagctc	ccagtcaagt	450
tggtgaatgt	ttgtgactca	ttcggaagt	atcatggggg	acctgcttat	500
attaggtgct	tggttgcaaa	caaacaaggc	agtcacgagg	ctgagctggg	550
aggatcactt	gagcctggga	agtggaggct	gcaataagcc	attattgtgt	600
tactgcactc	cagcctggca	cagaaaaaaaa	aaaaaaaaanac	aaactgagcc	650
agcaca					656

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Submission no : 11
Intron 10 : <..450
Exon 11 : 451..660
Intron 11 : 661..>

gatcacttct	aaagttaa	gtccatgga	aaacagtctc	atccacatct	50
ctttctggag	gccttccaag	cgtgctccat	gcagctctgt	tgcctgcccc	100
tgcacaggg	aatggaggct	ctgctttatc	ctgccctgtg	gtgtgactcc	150
cagaggcatc	agatgtggct	gggagtggga	gacatggaaa	attggctcct	200
gcaacagaga	actatcagcc	ttcccatcaa	ttggttactt	ctaattctgt	250
tatttttcag	gggcactgtc	ttctcataag	ctccatctat	gcaaaactaa	300
gccccatgggt	catgatgggt	ccctcaggcc	agaggcttgc	tggagagact	350
aatggatccc	ctggctaaaa	tctgtgcttg	ggctgcacat	tgggtaattt	400
cttctgaagg	aacagcctga	gcctgacatt	ctccatcttt	tccttggcag	450
GTTCTCCCTT	CGCCCGAGCC	AGCATTAAAA	GTGCCAAGCT	GGAGAACTCG	500
ACCTTTTTTC	ACAAAAAGGA	GAGGAGGATG	CGTTTCTACA	TCCGCCGCAT	550
GGTCAAAACT	CAGGCCTTCT	ACTGGACTGT	ACTCAGTTTG	GTAGCTCTCA	600
ACACGCTGTG	TGTTGCTATT	GTTCACTACA	ACCAGCCCGA	GTGGCTCTCC	650
GACTTCCTTT	gtgagtatca	cccagcccca	cccctgccaa	ctccctgate	700
cctccctcac	accctttttc	cacttctctt	tctctggtag	tatgtgtatc	750
ttctttgggc	ctcattgaat	ctgccctt			778

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Submission no : 12
Intron 11 : <..323
Exon 12 : 324..436
Intron 12 : 437..>

gatcacttgt	ggccaggagt	tcaagancag	ccagggcaac	atagtgagga	50
cccccatctc	cacattaaaa	attttaaaaa	gaaaaaagat	aagtcagaag	100
ttgggtgtgg	tgacacatgc	ctgtagttct	agcatgttgg	aggccaaatc	150
agggaaactg	tttgaggcca	ggagtttgaa	accagcctaa	cagcatagca	200
agacctcatc	tctacaaaaa	ataaaaaagt	taaaaatgat	aataaaaagga	250
aagtcagagc	cacctggaac	ccctaccctc	agcaagccta	acctcctctc	300
tgtttcctcc	ttctcccttc	tagACTATGC	AGAATTCATT	TTCTTAGGAC	350
TCTTTATGTC	CGAAATGTTT	ATAAAAATGT	ACGGGCTTGG	GACGCGGCCT	400
TACTTCCACT	CTTCCTTCAA	CTGCTTTGAC	TGTGGGgtaa	gtgctcttgt	450
ttctaagagt	tcattttctcc	agctccttgc	tggaatgaca	gatacctgga	500
cacattaaag	ggagaaaggt	aaagtcaccc	ctgaatatga	gagactcaga	550
tggatgcaga	aggaatgaga	aaacaatcca	aacactggca	aggatacagt	600
gtaccagaa	ccctcaacca	ccgcca			626

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Submission no : 13
 Intron 12 : <..545
 Exon 13 : 546..658
 Intron 13 : 659..821
 Exon 14 : 822..953
 Intron 14 : 954..>

gacnngncat	gcacaccagc	ctgggtgata	agagcaagac	tcctctcaaa	50
ataaatgaat	aaataaaaaat	aaataaataa	ataagaggcc	gggtgcagtg	100
gctcaatgct	ttggaaagt	gaggccaaca	gttggagaga	ccaaagcagg	150
aggatggctt	cagcccagaa	gtttgaggcc	mgcctgggca	atactagcga	200
gacactatct	ctataaaaaat	gttttaaaaat	tagccagatg	tggtggggca	250
cacctgtaat	cccagctact	caagaggctg	aggtgggagg	atcacttaag	300
cccaggagga	cagtgcctgca	gtgagctatg	attgcgcccc	ctgcactcca	350
gcctgggtga	cacagtgaga	cccggctctct	atagataaat	gaatggatga	400
atgaggggggt	caaggatcct	caccgggctt	ccatttggag	ggaggagttt	450
ggttgagttc	ttgcaagggt	ggtacctagg	aaatgcttgc	cagttctgga	500
gcccagacac	tgtccctgga	catgagacca	ggttctctgc	cctagGTTAT	550
CATTGGGAGC	ATCTTCGAGG	TCATCTGGGC	TGTCATAAAA	CCTGGCACAT	600
CCTTTGGAAT	CAGCGTGTTA	CGAGCCCTCA	GGTTATTGCG	TATTTTCAAA	650
GTCACAAAg	aagtctttgg	ggttcctgga	catttgtaca	gggggtgggg	700
atgggggaca	tggtggggcc	gcctccagaa	agttgggaaa	gtgagcctcg	750
tgtttcgagg	gctgactccg	gggcctgcct	wccccgcctg	gcctgagtc	800
tcgcctggsc	tctgtcggca	gGTACTGGGC	ATCTCTCAGA	AACCTGGTCG	850
TCTCTCTCCT	CAACTCCATG	AAGTCCATCA	TCAGCCTGTT	GTTTCTCCTT	900
TTCCTGTTCA	TTGTCGTCTT	CGCCCTTTTG	GGAATGCAAC	TCTTCGGCGG	950
CCAgtaagtc	cttcacagga	attcaa			976

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Submission no : 14
 Intron 14 : <..201
 Exon 15 : 202..274
 Intron 15 : 274..>

ccctccacgt	gcaggctgcc	ttcctcgtag	cccagacacc	catttgcggt	50
cacccaaatg	ggcagggccc	tgggtaccac	tcagggtttc	ctggggacag	100
agatgatgga	aacgttcgtt	tccttggaga	tgagatactg	agccacaccc	150
tcagagcacc	ccgggtgggg	ccaacgtgaa	atgtctgtgt	cctccctgca	200
gGTTTAATTT	CGATGAAGGG	ACTCCTCCCA	CCAACTTCGA	TACTTTTCCA	250
GCAGCAATAA	TGACGGTGTT	TCAGgtacag	cctccacctg	gccccacggg	300
ccaacacctc	tcagtgtcac	agatgaaagt	gcctgctcca	catccaaggg	350
gcttccctga	actcctcctt	ctctacctgg	ccttttcaca	ccactttgaa	400
acacagattt	tatggttatc	attattcaat	tatggtgagg	ccaacagatc	450
aggagatgaa	tgtcattgga	aagatagttt	gtggctgggc	acggtggctc	500
acacccataa	tcccagcact	ttggccaggt	acggtggctc	acacctgtaa	550
tcccaacgct	ttgggaagcc	caggtgggagg	atcacttga	gatcaggaat	600
tcgagaccag	cctggccaan	atggtgaaac	cccattctta	ctaaaaatac	650
aaaaattagc	cgggcgtggt	agcacatgcc	tgtaatccca	gctactcggg	700
agatgaggca	caagaattgc	ttgaacctgg	gaggcagagg	ttgcagtgag	750
ccaagatcgc	cccactgcac	tcmagcctgg	gcaacagagt	gagactccat	800
ctcaaaaaag	caaaaagaaaa	aaaaaaaccac	tttgggaggt	caagatggga	850
ggactacttg	aggccaggag	tttgagacaa	gtctgggcaa	catagtgaga	900
ctccgtctct	gcaaaaaaat	wataataata	attagctggg	catggtgata	950
catacctcct	agctactagg	gcagctgaag	tggaaggatt	gcttaagccc	1000
aggaggttga	ggctgcagta	agctacaatc	acaccactat	actccagcct	1050
gggcgagaga	gcaaagccct	gtctcaaaaa	cgaaaagaaa	gtttgttata	1100
ctcacagatc					1110

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Submission no : 15
 Intron 15 : <..524
 Exon 16 : 525..642
 Intron 16 : 643..795
 Exon 17 : 796..863
 Intron 17 : 864.>

gatcctccca	ccttggcctc	ccaaagtgtt	gggattacag	gcatgagcca	50
tggcatgcgg	tctcttcctg	ttcttataag	ggcactaata	ccatcatgaa	100
gtcccccatg	acctcatcta	accctagtta	cctctttaaag	gccccatctc	150
caaataccat	cccatcatag	gttagggctt	caactcatga	at ttggaggc	200
gggcacaatt	tagtccataa	caaatccccct	taatcacatc	aagtaagaca	250
gagttacagg	aggggtctgtg	actcctccag	ggteccattt	tcctagaagc	300
caggctaaga	gccccacgac	gcaggaacgg	ccctttctac	tcgcaaacaa	350
agagaaaagc	caaggagaag	ccaacacgga	gtctggctct	gcaaaccggg	400
caggattgtt	aaagacctcc	tgggctcggg	gatgggggtg	gcggattccg	450
gctccacagc	tgcattctcca	aggggcccgt	ggctgagagg	gggggttggt	500
gtgtgtttct	tcctccccctt	tcagATCCTG	ACGGGCGAAG	ACTGGAACGA	550
GGTCATGTAC	GACGGGATCA	AGTCTCAGGG	GGGCGTGCAG	GGCGGCATGG	600
TGTTCTCCAT	CTATTTTCATT	GTA CTGACGC	TCTTTGGGAA	CTgtatcctt	650
catggagaga	gagaagggga	caggcctgga	cctctggcag	aggagagggt	700
gcaggggctc	aagggagggt	actgagagac	ccagataccc	agggcccaag	750
tgggtgtccca	ccagtgggtg	cttttcctga	ctcagacatt	tgcagACACC	800
CTCCTGAATG	TGTTCTTGGC	CATCGCTGTG	GACAATCTGG	CCAACGCCCA	850
GGAGCTCACC	AAGgtggagg	cggtggggaga	atgtttctct	ggcaaagtta	900
cgacctgccc	atggcagatc	aagcactttt	ttggattaac	tgagccacag	950
gaaataacat	tttcaaatag	atkaaaaaga	tc		982

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Submission no : 16
Intron 17 : <...119
Exon 18 : 120..226
Intron 18 : 227..>

ccttggttct	gattgggtcga	aatatttcaa	atggtgcccc	tggtcagcaa	50
caggggtcaga	agtgagtccc	caaggcctag	ttcatgtttt	gtgaacaaaag	100
attccacgtg	cctttttcagG	ACGAGCAAGA	GGAAGAAGAA	GCAGCGAACC	150
AGAAACTTGC	CCTACAGAAA	GCCAAGGAGG	TGGCAGAAGT	GAGTCCTCTG	200
TCCGCGGCCA	ACATGTCTAT	AGCTGTgtaa	gtccccctaat	ccctgggatg	250
cta~cctggc	tcctgaacgt	gtccgaccac	tatccaggca	cagatttggg	300
aagcagtggg	ggtg				314

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Submission no : 17
 Intron 18 : <..209
 Exon 19 : 210..1019
 Intron 19 : 1020..>

gcccctagcc	aggtgggagc	catggagggt	tcttgagcag	aggaggctgg	50
gacctgactc	agatgctcac	agactcctag	cattcagggtg	gggagtagag	100
ggtggagagc	aggagtggga	ggctgagatg	tgggttggtt	cgcctgggtc	150
atccatccaa	gctacagtgc	ctagcaatgc	tctaagctcc	tgtgaccatg	200
ccactgcagG	AAAGAGCAAC	AGAAGAATCA	AAAGCCAGCC	AAGTCCGTGT	250
GGGAGCAGCG	GACCAGTGAG	ATGCGAAAAGC	AGAACTTGCT	GGCCAGCCGG	300
GAGGCCCTGT	ATAACGAAAT	GGACCCGGAC	GAGCGCTGGA	AGGCTGCCTA	350
CACGCGGCAC	CTGCGGCCAG	ACATGAAGAC	GCACTTGGAC	CGGCCGCTGG	400
TGGTGGACCC	GCAGGAGAAC	CGCAACAACA	ACACCAACAA	GAGCCGGGCG	450
GCCGAGCCCA	CCGTGGACCA	GCGCCTCGGC	CAGCAGCGCG	CCGAGGACTT	500
CCTCAGGAAA	CAGGCCCGCT	ACCACGATCG	GGCCCGGGAC	CCCAGCGGCT	550
CGGCGGGCCT	GGACGCACGG	AGGCCCTGGG	CGGGAAGCCA	GGAGGCCGAG	600
CTGAGCCGGG	AGGACCCCTA	CGGCCGCGAG	TCGGACCACC	ACGCCCCGGA	650
GGGCAGCCTG	GAGCAACCCG	GGTCTGTTGA	GGGCGAGGCC	GAGCGAGGCA	700
AGGCCGGGGA	CCCCACCCGG	AGGCACGTGC	ACCGGCAGGG	GGGCAGCAGG	750
GAGAGCCGCA	GCGGGTCCCC	GCGCACGGGC	GCGGACGGGG	AGCATCGACG	800
TCATCGCGCG	CACCGCAGGC	CCGGGGAGGA	GGGTCCGGAG	GACAAGGCGG	850
AGCGGAGGGC	GCGGCACCGC	GAGGGCAGCC	GGCCGGCCCCG	GGGCGGCGAG	900
GGCGAGGGCG	AGGGTCCCCG	CGGGGGCGAG	CGCAGGAGAA	GGCACCGGCA	950
TGGCGCTCCA	GCCACGTACG	AGGGGGACGC	GCGGAGGGAG	GACAAGGAGC	1000
GGAGGCATCG	GAGGAGGAAg	taagtggagg	tgacctcgaa	tccgcagaat	1050
gacggttaaca	ttaataatac	aacagccaaa	gtagcacgtg	ctgtgtattt	1100
gttataaaat	ata				1113

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Submission no : 18
Intron 19 : <..67
Exon 20 : 68..531
Intron 20 : 532..>

gtcctgaaac	tttgcctttt	aatcctaaat	cattgttggg	tctttttcat	50
tcacttgcect	tcctcagAGA	GAACCAGGGC	TCCGGGGTCC	CTGTGTCGGG	100
CCCCAACCTG	TCAACCACCC	GGCCAATCCA	GCAGGACCTG	GGCCGCCAAG	150
ACCCACCCCT	GGCAGAGGAT	ATTGACAACA	TGAAGAACAA	CAAGCTGGCC	200
ACCGCGGAGT	CGGCCGCTCC	CCACGGCAGC	CTTGGCCACG	CCGGCCTGCC	250
CCAGAGCCCA	GCCAAGATGG	GAAACAGCAC	CGACCCCGGC	CCCATGCTGG	300
CCATCCCTGC	CATGGCCACC	AACCCCCAGA	ACGCCGCCAG	CCGCCGGACG	350
CCCAACAACC	CGGGGAACCC	ATCCAATCCC	GGCCCCCCCA	AGACCCCGGA	400
GAATAGCCTT	ATCGTCACCA	ACCCAGCGG	CACCCAGACC	AATTCAGCTA	450
AGACTGCCAG	GAAACCCGAC	CACACCACAG	TGGACATCCC	CCCAGCCTGC	500
CCACCCCCCC	TCAACCACAC	CGTCGTACAA	Ggtgagaccc	tctgctctca	550
catcactggg	caggggacct	ggcgtcctgg	agccagaggt		590

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Submission no : 19
Intron 20 : <..75
Exon 21 : 76..217
Intron 21 : 218..>

ggagtacacc	gaggagttcc	cagagacttg	tgggaaattg	tggagggagc	50
cctgtgttgg	ttcttgtccc	aacagTGAAC	AAAAACGCCA	ACCCAGACCC	100
ACTGCCAAAA	AAAGAGGAAG	AGAAGAAGGA	GGAGGAGGAA	GAAGACGACC	150
GTGGGGGAAGA	CGGCCCTAAG	CCAATGCCTC	CCTATAGCTC	CATGTTCATC	200
CTGTCCACGA	CCAACCCgtg	agtatggccc	ccgagcagag	ggcagggggg	250
gctgggtctc	ccaccagggt	ggcggaannn	nnnnnnnnnn	nnnnnnnctc	300
ccaccagggg	ggcggaagtc	aggccagatt	agagggcaat		340

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Submission no : 20
Intron 21 : <..97
Exon 22 : 98..227
Intron 22 : 228..>

gatctcagta	gtggtaggta	acatgagatt	atggaagaaa	agggtttgtg	50
agcctgtggt	ctgagtggac	ctctgcacgc	ccatctgtct	ccaacagCCT	100
TCGCCGCCTG	TGCCATTACA	TCCTGAACCT	GCGCTACTTT	GAGATGTGCA	150
TCCTCATGGT	CATTGCCATG	AGCAGCATCG	CCCTGGCCGC	CGAGGACCCT	200
GTGCAGCCCA	ACGCACCTCG	GAACAACgtg	agtcccacag	agcacacccc	250
ttcctagect	ggctgctctg	cctcaggcca	ctttctcctg	catccaaaat	300
gctcataggt	agggtaggat	gttgggggtca	cccctaggca	tagcccttat	350
ggctgctggt	tgagagggga	agctctgatt	ccttggggat	gctcttggga	400
gcaagacatt	ccttgaggca	gtttctctgt	gagcctggtg	gggtggaggt	450
ggcccagagt	gactggggct	gaaaatt			477

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Submission no : 21
Intron 22 : <..33
Exon 23 : 34..93
Intron 23 : 94..>

gatccactgc	tctcttgctt	ttatccctta	cagGTGCTGC	GATACTTTGA	50
CTACGTTTTT	ACAGGCGTCT	TTACCTTTGA	GATGGTGATC	AAGgtgagtg	100
cagattataa	gtgagaacac	acggtaattt	ttttttttta	gcaagtgcag	150
ggctgggcac	agtggatc				168

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Submission no : 22
Intron 23 : <..232
Exon 24 : 233..339
Intron 24 : 340..>

gatctaagag	ccggcaagcc	agagctggct	tccatcaggc	aaaggggggc	50
cgcctcatgg	ggcaggggct	ccccactcct	ccctgggagt	cctctggcca	100
ctgcccaccc	ctgcaagatg	aggtggcctc	attggcttcc	ctgcctctcc	150
ccgagaggct	agagagtggg	tggcagcacc	ccaggggtggg	gatcaggtgg	200
gggttctgag	caccctctct	tctccccccac	agATGATTGA	CCTGGGGCTC	250
GTCCTGCATC	AGGGTGCCTA	CTTCCGTGAC	CTCTGGAATA	TTCTCGACTT	300
CATAGTGGTC	AGTGGGGCCC	TGGTAGCCTT	TGCCTTCACg	taagtctctt	350
cgcaagggtt	tcctcttg				368

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Submission no : 23
Intron 25 : <..244
Exon 25 : 245..344
Intron 24 : 345..>

gatcttaacc	ccaagacact	tcatctaaag	gaaaaaactgc	cataatacac	50
agattatattt	aggtcagctc	acttttactgc	catctgctgg	gaagttgtaa	100
taatacaaat	atccatacac	gatggctagg	atgttatcag	cacctccttt	150
aatgtgttgt	ccttgagcag	tgtacaacct	gctcagctgt	acatgataac	200
cctgacagtc	ccccccaccg	cacccccacca	tctcccaatc	tcacCTTGAG	250
CTTTGGCAGC	CGCTTGATGG	TTTAAAGAGG	TCGTAGCACC	CGGAGGACTC	300
GGAGGGATTT	AATCGTGTTG	ATGTCTTTTC	CTTTGCTATT	GCCActgtgg	350
aggaatgttt	agggtgggaag	aagggaagag	aggaagcaga	ggtcagggtg	400
ggtagggggc	agcccacagc	tccatgggac	cctacccttc	ccaggcctag	450
aagtctgggg	tgagcttggc	acaagcctgc	cctttcctgg	tgaagagtgg	500
tccattttac	cctgt				515

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Submission no : 24
Intron 25 : <..67
Exon 26 : 68..228
Intron 26 : 229..>

ggccactgga	ggcagaaggt	tggcaggtcc	ccagcccctc	atgctctctg	50
tcaactccac	cccacagGCT	GTGTTTGACT	GTGTGGTGAA	CTCACTTAAA	100
AACGTCTTCA	ACATCCTCAT	CGTCTACATG	CTATTCATGT	TCATCTTCGC	150
CGTGGTGGCT	GTGCAGCTCT	TCAAGGGGAA	ATTCTTCCAC	TGCACTGACG	200
AGTCCAAAGA	GTTTGAGAAA	GATTGTCGgt	gggtctccgc	tttccagcac	250
attcccattg	gaaccagcag	gtgggcaggg	gggaagtggc	tagaggcatt	300
ggccacttgg	gctcagagac	tggagaagtg	atgagccttg	gaagtgactc	350
agttgcaacc	agcttggatc	aagggtagaa	agaaaaccgg	ttttagaatt	400
tgagtc					406

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Submission no : 25
 Intron 27 : <..177
 Exon 27 : 178..315
 Intron 26 : 316..>
 Remark : reversed direction!

gatctcaaac	tcttggcctc	aagtgatata	tctgccttgg	cctcctaaag	50
tggtgggatt	acaggcgtga	gcaccatgcc	cggcctccaa	gacctttctt	100
attgctaagc	tctcaggccc	tttatcctcc	tgctccccag	ggctcctcct	150
ggatagattt	ccagtcgggc	cacttacTGT	GGCCAGCCTT	CTCCCGTGGA	200
CACGGTGAAG	AGGGTCAGCA	GAGCCCACAG	CACATTGTCTG	TAATGGAATT	250
CATACTTCTT	CCACTCCCGG	TCTCGCGCCT	TCACCTCATT	CTTCTCGTAG	300
AGGAGGTATT	TGCCTctgcc	acagagagtg	gggactgtta	gtaaatggga	350
aagaggggct	gtcttgcaact	tgtctttggt	tatcagagac	agggggaggg	400
aaaggaagag	tggteccacca	ncctagactg	cttggggaagc	agtgaactcc	450
catcctgcca	ccatgtgttc	ctgtgcttca	taggggatgn	cgtgtgcaat	500
ctacttttna	ggataa				516

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Submission no : 26
Intron 27 : <..84
Exon 28 : 85..276
Intron 28 : 277..>

accttcctca	tcacccttgg	gtccctgtct	ctctccttcc	tgcccccttcc	50
ctctccctgc	cccattcctt	gcagGGTCCT	CAAGCATTCG	GTGGACGCCA	100
CCTTTGAGAA	CCAGGGCCCC	AGCCCCGGGT	ACCGCATGGA	GATGTCCATT	150
TTCTACGTCG	TCTACTTTGT	GGTGTTCCCC	TTCTTCTTTG	TCAATATCTT	200
TGTGGCCTTG	ATCATCATCA	CCTTCCAGGA	GCAAGGGGAC	AAGATGATGG	250
AGGAATACAG	CCTGGAGAAA	AATGAGgtgc	cacttccaat	tccatctgtc	300
ctttaaaaaac	tggggacaca	cacaaaacttt	aaaacacaca	caacacccag	350
gaaccccttt	ctaggggtac	ctggggggagg	gaacagaagc	attgtcccaa	400
ccgaatccag	tcttcagggc	agcccttcat	ggagtttcag	aggaaacaca	450
tcatatagtg	tatgtatcag	tcagtttaga	ctaggttat		489

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Submission no : 27
Intron 28 : <..253
Exon 29 : 254..418
Intron 29 : 419..>

tagcccatgc	aanaatgggg	aaatgncagt	gcaagttttg	gcagttgntg	50
acatctcaag	caactgtanc	tgttgggata	agaaagcaat	ggtgagaagg	100
aanagaganc	ccaggaatcc	tggctggggg	caananaggc	agagactcaa	150
gcagaagcac	ttgagaaccg	cgacgagtta	gacagagggg	gcccgggtga	200
cagccacctt	cctcctgcct	ctgccgetct	caccactggc	ctctctcccg	250 50
cagAGGGCCT	GCATTGATTT	CGCCATCAGT	GCCAAGCCGC	TGACCCGACA	300
CATGCCGCAG	AACAAGCAGA	GCTTCCAGTA	CCGCATGTGG	CAGTTCGTGG	350
TGTCTCCGCC	TTTCGAGTAC	ACGATCATGG	CCATGATCGC	CCTCAACACC	400
ATCGTGCTTA	TGATGAAGgt	aagtgcccc	caccagcccc	cagcactant	450
taacccccac	ctcgttcctg	cctctaccct	gataaaatga	aaccatttgc	500
agattttccca	ga				512

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Submission no : 28
Intron 29 : 156
Exon 30 : 157..267
Intron 30 : 268..>

gggtcttttc	tgaactgtgc	ctcctaccag	tgagggtgct	cagaccttgc	50
ctggggctgg	agtgttgct	ggagaacagc	catgaagctg	acctccccac	100
ttcccacttc	ccaccctgc	tcgctgaccc	ctgctactcc	tgcttctttc	150
ccctagTTCT	ATGGGGCTTC	TGTGGCTTAT	GAAAAATGCC	TGCGGGTGTT	200
CAACATCGCC	TTCACCTCCC	TCTTCTCTCT	GGAATGTGTG	CTGAAAGCCA	250
TGGCTTTTGG	GATTCTGgta	agtaccacct	tggggctaca	gctatgggct	300
tgganaanc	ccaaggggga	acaatgggtc	ctggatgatg	gtctcccaac	350
gtggcccca	gaacccaac	ctcaaggggtg	gcttcagtat	cctgcccagt	400
ggccacagat	c				411

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Submission no : 29
Intron 30 : <..115
Exon 31 : 116..199
Intron 31 : 200..>

ctgtcccggg	cactccgctg	atgggcaact	gtgcctctaa	catgcaccgg	50
ccagcctagg	gggccgggaa	ccaagccctc	tggtggcatc	tctgtcttgt	100
gggtcccat	tctagAATTA	TTTCCGCGAT	GCCTGGAACA	TCTTCGACTT	150
TGTGACTGTT	CTGGGCAGCA	TCACCGATAT	CCTCGTGA	GAGTTTGGGg	200
taagtctccc	tccagcttct	ctctgggtga	ctctgggctg	gacgaggcag	250
gcggcagggg	gcgggggagc	ggccccagag	gcagtgtgtc	ccggaagcca	300
tagctgcttg	agccagcact	tggccatgac	cagagagggg	gaactggggc	350
cccggggaca	agggcagccc	ctcaggaggg	cattgtgggg	agatgggggt	400
aacaaagctt	ggctgtaggg				420

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Submission no : 30
Intron 31 : <..148
Exon 32 : 149..265
Intron 32 : 266..>

ttaatagtg	cttctctctc	cctccttatt	tggggtctgg	cttgcttttt	50
tcctggttg	tggcttcatg	taggggcctc	tgtgagtgg	gacagctctg	100
agcctttgg	gtgggtggat	ggtcacccct	cttctctcat	ctccccagAA	150
TAACTTCATC	AACCTGAGCT	TTCTCCGCCT	CTTCCGAGCT	GCCCGGCTCA	200
TCAAACCTTCT	CCGTCAGGGT	TACACCATCC	GCATTCTTCT	CTGGACCTTT	250
GTGCAGTCCT	TCAAGgtgag	tcctcgtccc	tgctgctggc	ccaggggctg	300
agaagacagg	tgaccctcat	gctctggctg	aatgtagaag	tc	342

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Submission no : 31
 Intron 32 : <..156
 Exon 33 : 157..222
 Intron 33 : 223..394
 Exon 34 : 395..509
 Intron 34 : 510..>

cccccaagaa	gaatgcccac	caagccctgg	aaggactctg	gcacgtggca	50
tatgyccacc	caacccagtg	gggcagagca	ctgggacaag	ggaggaagac	100
tgcagtgcgg	ctgaggggacc	cccagcactc	ttcttcattg	ccttttttcc	150
caccagGCCC	TGCCTTATGT	CTGTCTGCTG	ATCGCCATGC	TCTTCTTCAT	200
CTATGCCATC	ATTGGGATGC	AGgtgagtgt	cgtgtcccta	aggttcccag	250
agcctcccaa	ggaggggcagc	cacccttaga	aaggggtggg	tcagaggagc	300
ctggttcaca	gaagcagcca	tggaggttga	gctgggtttc	ccagaagcca	350
ctggaggaat	ggcagccccct	ggtcgtcacc	cwmcaattcc	acagGTGTTT	400
GGTAACATTG	GCATCGACGT	GGAGGACGAG	GACAGTGATG	AAGATGAGTT	450
CCAAATCACT	GAGCACAATA	ACTTCCGGAC	CTTCTTCCAG	GCCCTCATGC	500
TCTCTTCCGg	tcagaagggg	acctgctctg	ataatnctgt	ttccgtgggg	550
tgggggtgcc					559

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Submission no : 32
Intron 34 : <..94
Exon 35 : 95..245
Intron 35 : 246..>
Sequence : 316

tcagagccat	gctcactgtg	tgctccactc	ctgaggaggc	gttggtacca	50
gtcagggctg	ggtgtccgag	tctctgattt	ctccctgtcc	tcagGAGTGC	100
CACCGGGGAA	GCTTGGCACA	ACATCATGCT	TTCCTGCCTC	AGCGGGAAAC	150
CGTGTGATAA	GAACCTCTGGC	ATCCTGACTC	GAGAGTGTGG	CAATGAATTT	200
GCTTATTTTT	ACTTTGTTTC	CTTCATCTTC	CTCTGCTCGT	TTCTGgtgag	250
tctgtggaca	ctgtgagggc	cgtctgggct	ccctaagcct	ggcttccttt	300
cagggagtgg	ttctgt				316

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Submission no : 33
Intron 35 : <..211
Exon 36 : 212..339
Intron 36 : 340..>

gtgtagtgag	aactcacctc	tccattcccc	agtctctttc	tgtctctgtc	50
tcatttcctt	tccccatctt	ctctctatcc	ctctctccat	ctggggcctc	100
tgtgtctgtc	tttgggtctg	tctgtccgtc	tgactgtctg	tatccttctc	150
acttcactca	ttcattccct	cggctctctg	cccattctct	cttgggtccc	200
ggtccccaca	gATGCTGAAT	CTCTTTGTCTG	CCGTCATCAT	GGACAACTTT	250
GAGTACCTCA	CCCGAGACTC	CTCCATCCTG	GSCCCCCACC	ACCTGGATGA	300
GTACGTGCGT	GTCTGGGCCG	AGTATGACCC	CGCAGCTTGg	taagaagtca	350
ccccgaatcc	tccagccaca	atactcacct	ctccctggaa	ctggaacacg	400
ggctaggcta	ggnccccaga	ctctggagca	ctgaactcct	ggggctccta	450
gcaggggtct	cacaggttca	gtcaggagag	aagatataag	aatcatcacc	500
cttgcatacc	ccagattaaa	cacgtagggt	gccaacctgc	ccaaacctg	550
gaggactttc	tgggaaatga	ggagggcgct	aaccatgaga	tgtctgaaga	600
gcccctctct	cctacgagtc	tctcctgtct	ctcactgtga	agtctccaga	650
tgggtgaggat	cgattagcca	ggctccagga	gaaaccaaca	gact	694

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Submission no : 34
Intron 36 : <..213
Exon 37 : 214..310
Intron 37 : 311..>

aagggaggtg	cctgcagtc	cgaactcgac	tgacatccta	cacccctggg	50
tctccccagt	gtctgggaat	gtactgggaa	ttcacttgtc	cccagtctct	100
cccactcctt	gaagccaggg	acaccccgac	ctcgggcatc	atgacctcgt	150
tgtgtgcca	gggagcccgt	gtgaacccat	tgccctgcact	aacccctttt	200
cttctccttt	cagCGGTCGG	ATTCATTATA	AGGATATGTA	CAGTTTATTA	250
CGAGTAATAT	CTCCCCCTCT	CGGCTTAGGC	AAGAAATGTC	CTCATAGGGT	300
TGCTTGCAAG	gtttgacttc	cactaaaacc	tgctagcatc	catggaatga	350
gtgtggccttg	gggttcttca	atatatatat	ttcatatata	tatatatata	400
tatctctctc	tctctaaaaa	aacagagcca	tctctctttc	ttgcattaaa	450
ctagaaaact	ctcttagcca	acag			474

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Submission no : 35
Intron 37 : <...82
Exon 38 : 83..188
Intron 38 : 189...>

cctgggtagg	ggcgggcgcg	gctcacggga	gacccaggag	ggatgcctgg	50
gaatgactgc	gcttgccttg	ggttttctgt	agCGGCTTCT	GCGGATGGAC	100
CTGCCCCGTCG	CAGATGACAA	CACCGTCCAC	TTCAATTCCA	CCCTCATGGC	150
TCTGATCCGC	ACAGCCCTGG	ACATCAAGAT	TGCCAAGGgt	aaggaagggga	200
caggggcggg	cacagacagg	cgtgacaggg	tggaaactggg	gatctcctcc	250
ctacccccaaa	ctagaggatc	tgctgtcacc	acccggatct	tcattcactc	300
ttccattcat	tcgttcaca	ggnnntttttg	gnnnnttggnn	ntttggtgtt	350
tttttttttt	ttttgagaca	gagtccttgct	ctggttgcca	ggcagcagtg	400
cggtgacatg	atc				413

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Submission no : 36
Intron 38 : <..96
Exon 39 : 97..204
Intron 39 : 205..369
Exon 40 : 370..470
Intron 40 : 471..>

gggtctcggt	ctcgggagcc	tatggctttg	cagctgaccc	agagtccagc	50
tgacacccag	gcaggcagtc	agggctctgtc	tacaccccca	ttgcagGAGG	100
AGCCGACAAA	CAGCAGATGG	ACGCTGAGCT	GCAGGAAGGAG	ATGATGGCGA	150
TTTGGCCCCA	TCTGTCCCAG	AAGACGCTAG	ACCTGCTGGT	CACACCTCAC	200
AAGTgtaaga	gctgagccca	gccctgggat	ccaatccacc	aggacagatg	250
gag_gggagg	gaaaggggag	gcctggggag	agtgttggct	gggctgggtat	300
acacagggac	ccaggacaag	gtccccaag	angcctgccc	ttggtgagct	350
caccgtgtgt	gtccccagc	CACGGACCTC	ACCGTGGGGA	AGATCTACGC	400
AGCCATGATG	ATCATGGAGT	ACTACCGGCA	GAGCAAGGCC	AAGAAGCTGC	450
AGGCCATGCG	CGAGGAGCAG	gtgcgctgtt	cgccgctctg	gggacatctg	500
ggctggggac	agtggcttgc	atgtcaccac	gggaaccaac	tggaatatga	550
gggtggctga	gcccagggc	aggtccctga	aaagttagggg	ctggtgcaca	600
gcagctcaca	cctgcaatct	cagtgcctttg	agaggc		636

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Submission no : 37
 Intron 40 : <..407
 Exon 41 : 408..517
 Intron 41 : 518..625
 Exon 42 : 626..764
 Intron 42 : 765..>
 Sequence : 829

gatcttcagg	gccatgggag	ctgcaggaag	gactctggct	ttttcccca	50
gcaagtggga	gccatggagg	gttctaagca	aaggagggat	aggacctgac	100
tcaagtgctc	atgggcgccc	tctggtggct	cttgtggaac	agtggggttg	150
aaggtaggag	cgggagacct	gggagaaggt	gcctgcagtg	agagatgagg	200
acgcgggacc	aggctggggc	tatgacttgg	gtggaggagt	gagaagtggg	250
ccagttctgc	gtggaattgg	aagggtctag	atggatgaga	cctgagagag	300
tgtgtgtgtg	tgtgtgtgtg	tatactgggg	atgtcgcaat	gccttctggg	350
taccacegtc	caccacccca	cccttgtcca	cacactgctc	tctgccccat	400
tccccagGAC	CGGACACCCC	TCATGTTCCA	GCGCATGGAG	CCCCCGTCCC	450
CAACGCAGGA	AGGGGGACCT	GGCCAGAACG	CCCTCCCCCTC	CACCCAGCTG	500
GACCCAGGAG	GAGCCCTgtg	agtgtcaccc	ctgccaggga	ggtggagtgt	550
gggggtgccc	tgggtccccac	gttctggaag	ctgcccgaagc	gcccactgct	600
accccggect	ctgtccccca	tgcaagATGG	CTCACGAAAG	CGGCCTCAAG	650
GAGAGCCCGT	CCTGGGTGAC	CCAGCGTGCC	CAGGAGATGT	TCCAGAAGAC	700
GGGCACATGG	AGTCCGGAAC	AAGGCCCCCC	TACCGACATG	CCCAACAGCC	750
AGCCTAACTC	TCAGgtgcct	ctgtccccca	actccccaat	ggctcccagg	800
gcccgggtgg	ttgcggtgga	aggaaccat			829

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Submission no : 38
Intron 42 : <..219
Exon 43 : 220..333
Intron 43 : 334..>

tcactgcaac	ctccaccttc	cagtctcaag	tgattcctcc	tgccctcagcc	50
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tttgtatttt	tagtagagac	ggggtttcac	aatgttggtc	aggctgggtct	150
cgaactgctg	nccattgtga	tctggaggtc	aggccccaga	gtcatctctgg	200
ctttgccatt	cgtccgcagT	CCGTGGAGAT	GCGAGAGATG	GGCAGAGATG	250
GCTACTCCGA	CAGCGAGCAC	TACCTCCCCA	TGGAAGGCCA	GGGCCGGGCT	300
GCCTCCATGC	CCCGCCTCCC	TGCAGAGAAC	CAGgtgaggg	ctttcaccac	350
tgccctgggg	ctggacccct	cactctgcac	tgggtagggc	caggcccccc	400
cacaagcagc	ccagtgcac	ccctcctgcc	ggactcaggc	ctgggtaggg	450
actccttcag	tctctgaagc	agtctgcagg	ccccaccac	cacctgggtca	500
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ggccaggagt	ttgagaccag	gttgggcaac	atagcaaaac	cctgtctctt	750
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a					801

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Submission no : 39
Intron 43 : <..83
Exon 44 : 84..119
Intron 44 : 120..>
Sequence : 329

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tccggccccc	tcttcctccc	caccccnct	ccagggagtc	ccgtaatctc	200
taccgggtccc	cggacccac	cctttctttg	gcaatcgac	cctctcccct	250
ccatggagcc	caatccttgt	gtgtggtgtc	ctgtgtgtgc	cctgacccat	300
aagcctggtg	gggcggccat	ccccatcct			329

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Submission no : 40
Intron 44 : <..166
Exon 45 : 167..353
Intron 45 : 354..>

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aggaaggacc	gggaccactt	ggtgctagct	gctgacccca	gcccaccggc	150
ctgtccctc	ccccagACCA	TCTCAGACAC	CAGCCCCATG	AAGCGTTCAG	200
CCTCCGTGCT	GGGCCCCAAG	GCCCGACGCC	TGGACGATTA	CTCGCTGGAG	250
CGGGTCCCGC	CCGAGGAGAA	CCAGCGGCAC	CACCAGCGGC	GCCGCGACCG	300
CAGCCACCGC	GCCTCTGAGC	GCTCCCTGGG	CCGCTACACC	GATGTGGACA	350
CAGgtgggca	gccctgtggt	gctcagggac	aagcagaaca	gaggagagga	400
gaggggagga	gaaggcaggg	cggaggagac	actaagggaag	aagaaagggg	450
gaggcctcca	tggagagggg	acagagcggg	ccaggcagcg	gctgcaggaa	500
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gatc					554

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Submission no : 41
Intron 45 : <..31
Exon 46 : 32..285
Intron 46 : 286..>

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GGGGCCGGCC	CAAGGATCGG	AAGCATCGAC	AGCACCACCA	CCACCACCAC	150
CACCACCACC	ATCCCCCGCC	CCCCGACAAG	GACCGCTATG	CCCAGGAACG	200
GCCGGACCAC	GGCCGGGCAC	GGGCTCGGGA	CCAGCGCTGG	TCCCGCTCGC	250
CCAGCGAGGG	CCGAGAGCAC	ATGGCGCACC	GCCAGgtggg	tgcggctgca	300
agtgacccca	ggctgggctc	ggccggggagg	cggggaggag	agaaggggat	350
accccatcca	acagccactc	taggcaaagg	tccccggatc	ccggctgtga	400
ccacctccca	tcctgcccc	aagccaccgg	ggtgccccggc	ggccggagcg	450
gagcacggat	c				461

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Submission no : 42
Intron 46 : <..279
Exon 47 : 280..>
Stop codon : 280..282
UTR 3' : 283..>

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ttttttccatt	caatgtcctg	tgteccccct	ctcctcctcc	tcctcctcct	150
ccccctcctc	cctctcctcc	cggeccccct	cccttcgctc	ccctcatctt	200
cctcccaatc	ccgtgtctcc	tttgattttg	ttgtatcttt	ttttttgatt	250
tccttttgtt	caattttcgt	gtagggcagT	AGTTCCGTAA	GTGGAAGCCC	300
AGCCCCCTCA	ACATCTGGTA	CCAGCACTCC	GCGGCGGGGC	CGCCGCCAGC	350
TCCCCCAGAC	CCCCTCCACC	CCCCGGCCAC	ACGTGTCCTA	TTCCCCCTGTG	400
ATCCGTAAGG	CCGGCGGCTC	GGGGCCCCCG	CAGCAGCAGC	AGCAGCAGCA	450
GCAGCAGCAG	CAGGCGGTGG	CCAGGCCGGC	CGGGCGGGCA	CCAGCGGGCC	500
TCGGAGGTAC	CCAGGCCCCA	CGGCCGAGCC	TCTGGCCGGA	GATCGGCGCC	550
CACGGGGGGC	CACAGCAGCG	GCCGCACGCC	CAGGATGGAG	AGGCGGGTCC	600
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GGCTGGCGGC	AGTC				664

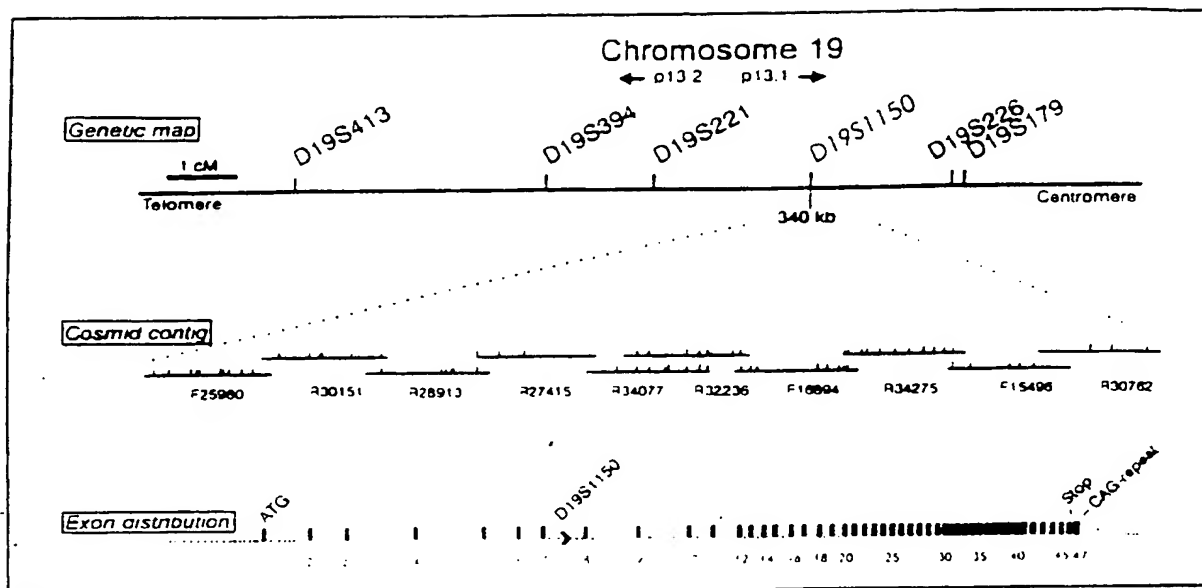


Fig. 2

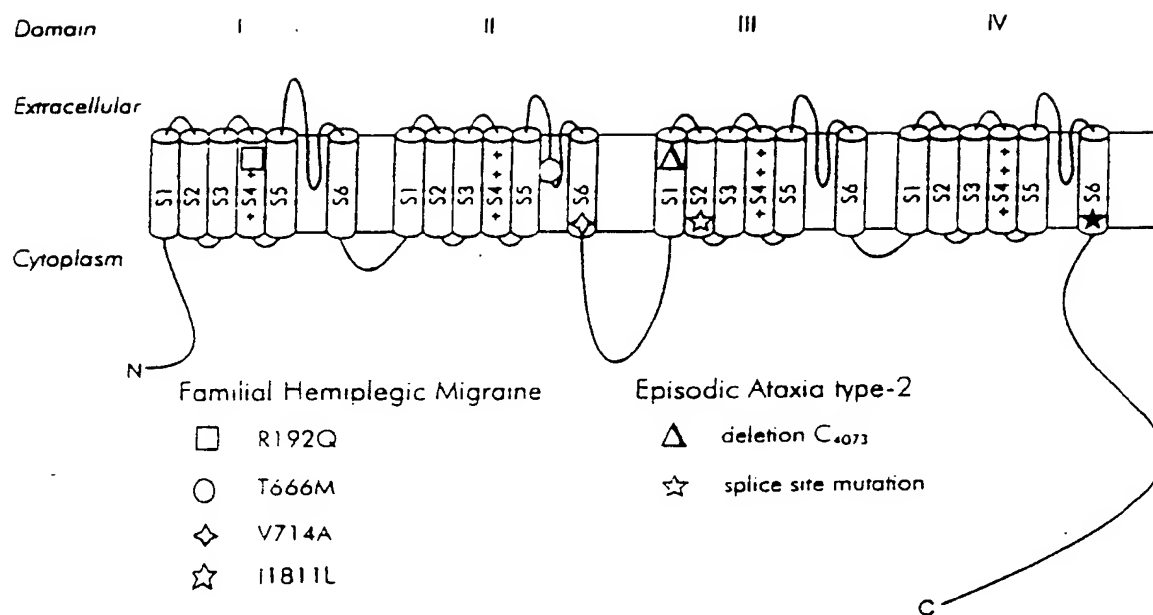


Fig. 3

Figure 4

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A G V V V G S G G G R G A G G S R Q G G 40
cagccccggggcgcaaaggatgtacaagcagtcgaatggcgagagagcgcgaccatggca 180
Q P G A Q R M Y K Q S M A Q R A R T M A 60
ctctacaacccccatccccgtccgacagaactgcctcacggttaaccgggtctctctctc 240
L Y N P I P V R Q N C L T V N R S L F L 80
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F S E D N V V R K Y A K K I T E W P P F 100
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E Y M I L A T I I A N C I V L A L E Q H 120
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L P D D D K T P M S E R L D D T E P Y F 140
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I G I F C F E A G I K I I A L G F A F H 160
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K G S Y L R N G W N V M D F V V V L T G 180
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I L A T V G T E F D L R T L R A V R V L 200
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K A M I P L L Q I G L L L F F A I L I F 240
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T D L L Y N S N D A S G N T W N W L Y F 340
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 I L L T F V Q S F K A L P Y V C L L I 1700
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